



REVIEW

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# Immune microenvironment dynamics in pregnant patients with concomitant autoimmune diseases: mechanisms, challenges, and clinical significance

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## Abstract

Throughout pregnancy, the immune microenvironment undergoes dynamic changes in patients with concomitant autoimmune diseases (AIDs). These alterations not only affect disease activity and clinical manifestations, but also play a pivotal role in sustaining maternal-fetal immune tolerance and pregnancy outcomes. Extensive preclinical studies have elucidated the mechanisms of immune regulation in normal pregnancy, whereas the dynamic immune changes in pregnancies complicated by AIDs remain poorly understood. Recent studies have revealed significant variations in immune responses to pregnancy among individuals with AIDs, which may contribute to distinct patterns of AIDs flares during this period. Despite substantial progress in immunology and reproductive medicine, comprehensive reviews addressing the dynamic changes in the immune microenvironment during pregnancy in the context of AIDs are lacking. In this review, we summarize existing knowledge and incorporate recent multidisciplinary findings, focusing on the dynamic changes in systemic immune adaptation and maternal-fetal immune interactions in the context of AIDs during pregnancy. We emphasize the clinical significance of these immune dynamics for optimizing management and therapeutic strategies. Additionally, we propose new perspectives and provide recommendations to guide future research and the development of personalized treatment approaches.

**Key words** Autoimmune diseases (AIDs), Pregnancy, Immune microenvironment, Maternal-fetal immune adaptation, Clinical management

## Background

Autoimmune diseases (AIDs) are characterized by disrupted immune homeostasis, a disorder that is particularly evident in women of reproductive age [1]. Accumulating evidence indicates that pregnant women with AIDs face a significantly increased risk of adverse pregnancy outcomes (APOs) [2]. During pregnancy, both systemic immune changes and localized immune adaptations at the maternal-fetal interface, especially those regulating the placental microenvironment, are essential for sustaining maternal-fetal immune tolerance and promoting favorable pregnancy outcomes [3]. In patients with AIDs, particularly autoimmune rheumatic diseases,

gestation is associated with dynamic shifts in disease activity. Importantly, gestational immune adaptation is heterogeneous across different autoimmune disorders, which may underlie distinct, disease-specific patterns of flares and remission [4,5]. While significant advances have been achieved in elucidating normal pregnancy immunoregulatory processes [6,7], essential scientific issues remain, especially concerning the intrinsic differences in gestational immune modulation between AID-affected pregnancies and normal pregnancies, as well as disease-specific immune dynamics across various autoimmune disorders. Within this context, we systematically integrate recent multidisciplinary evidence to delineate the sequential and dynamic alterations of maternal-fetal immune regulation across pregnancy stages, clarify the interplay between disease-specific immune profiles and pregnancy immune adaptation, and underscore the significance of clinical management and treatment strategies in AIDs during pregnancy.

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## Changes in the immune microenvironment during pregnancy

### Systemic immune microenvironment

During normal pregnancy, the maternal immune system undergoes finely tuned adaptive changes aimed at maintaining fetal tolerance. The complement system exhibits a dynamic balance, with increased activation counterbalanced by elevated inhibitory factors, providing anti-infective protection while avoiding fetal injury [8]. Concomitantly, granulocyte dynamics shift markedly, with neutrophil counts progressively increasing throughout pregnancy, accompanied by elevated levels of hematopoietic factors such as granulocyte colony-stimulating factor (G-CSF) and granulocyte-macrophage colony-stimulating factor (GM-CSF) [9,10], yet their function can be altered. Although resting neutrophils show increased activity, upon activation, their oxidative burst and reactive oxygen species production capacity decrease [11]. These adaptations are likely crucial for sustaining immune tolerance and preventing harmful immune reactions directed against the fetus.

In parallel, monocytes exhibit dynamic changes throughout pregnancy. From early pregnancy, circulating monocyte counts progressively increase, marked by an increase in intermediate monocytes and a decrease in classical subsets [12,13]. Concurrently, maternal monocytes show a shift toward proinflammatory subsets (e.g., increased IL-6<sup>+</sup>CD14<sup>+</sup> or MIP-1 $\alpha$ <sup>+</sup>CD14<sup>+</sup> cells) together with increased reactive oxygen species production, indicating enhanced proinflammatory potential [14]. Conversely, in late pregnancy, monocytes adopt an anti-inflammatory phenotype, characterized by reduced cytokine production upon lipopolysaccharide stimulation compared with that in nonpregnant women, indicating a state of immune tolerance [15,16]. This late-gestational monocyte hyporesponsiveness is conceptually reminiscent of sepsis-associated immunoparalysis, in which monocytes suppress proinflammatory responses to limit tissue damage [17-19]. This monocyte tolerance in late pregnancy likely serves as a critical immunoregulatory mechanism to preserve maternal immune homeostasis and prevent excessive immune activation [15]. In general, monocytes during pregnancy display characteristics of chronic low-grade inflammation and retain the capacity to restore immune function when appropriately stimulated, thus supporting immune balance between the mother and fetus.

Natural killer (NK) cells, which are vital components of the innate immune system, exhibit remarkable plasticity in their activity during pregnancy, enabling them to adapt to distinct local and systemic environments [20,21]. In the peripheral

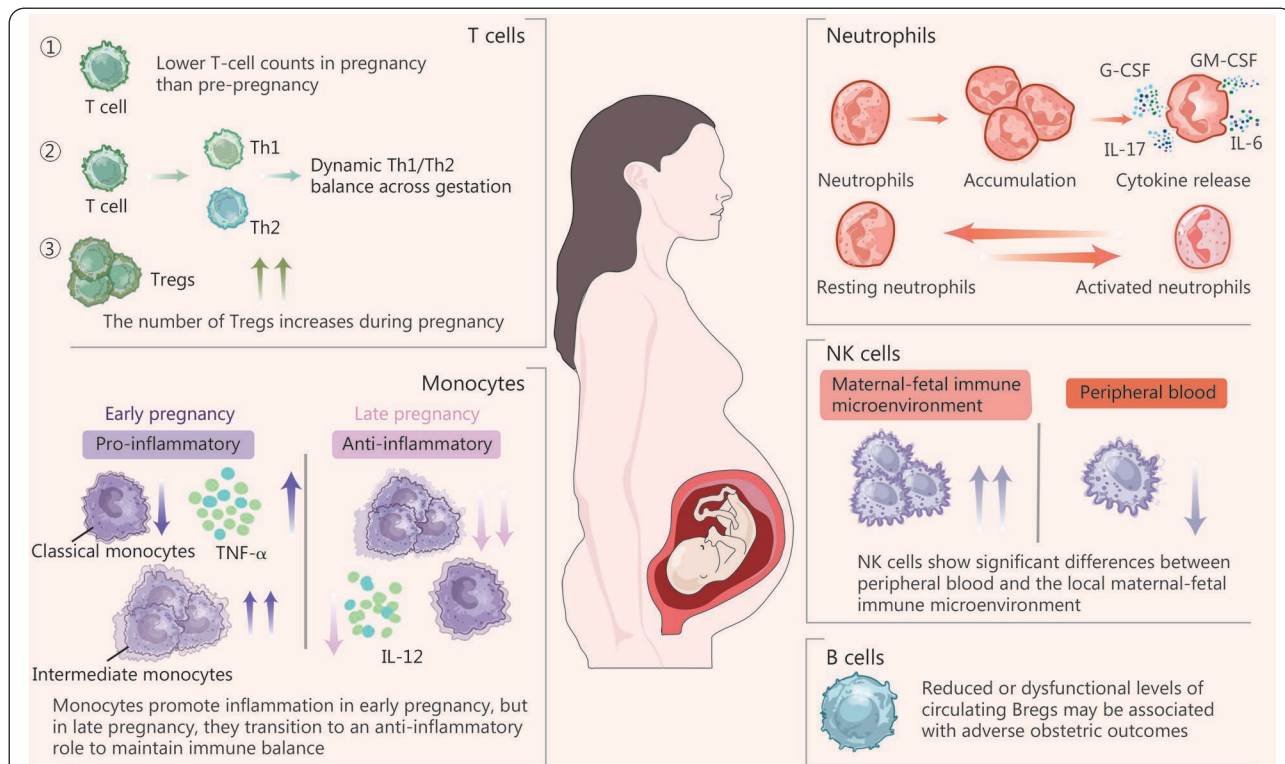
blood, the numbers and activity of NK cells are generally reduced, which helps prevent fetal rejection [22]. In contrast, a notable increase occurs in both the quantity and activity of NK cells within the placental microenvironment [23]. This functional divergence between placental and circulating NK cells highlights the specificity and complexity of placental immune regulation.

T cells undergo key systemic adaptations during pregnancy to maintain maternal immune homeostasis. While total lymphocyte and T cell percentages remain stable, overall T cell numbers decline compared with those in the pre-pregnancy state [24]. Regarding subset distribution, CD4<sup>+</sup> and CD8<sup>+</sup> T cells within the  $\alpha\beta$  T cell population tend to decrease in early and late pregnancy, likely due to the markedly increased estrogen and progesterone levels characteristic of pregnancy [25]. Sex hormones modulate thymic function, thereby shaping T cell development and regulating peripheral T cell counts, which is essential for sustaining maternal immune balance [26]. Functionally, a dynamic shift occurs, a T helper (Th)1-dominant profile supports implantation in early stages, followed by Th2-mediated immune tolerance during mid-to-late pregnancy, with a re-establishment of Th1 predominance towards term to initiate labor [27,28]. In addition, regulatory T cells (Tregs) play a pivotal role in maternal-fetal immune tolerance. During early pregnancy, peripheral Treg proportions markedly rise, enabling the suppression of autologous T cell proliferation and facilitating immune tolerance [29]. However, reports on Treg dynamics vary, likely due to inconsistent definitions, as both the CD25 and forkhead box P3 (FOXP3) markers are shared by activated T cells [30]. Accurate distinction is essential for assessing Treg fluctuations during pregnancy. Additionally,  $\gamma\delta$  T cells exhibit unique spatiotemporal distribution patterns. During the early phase of a normal pregnancy,  $\gamma\delta$  T cells increase in both the peripheral blood and decidua, predominantly comprising the V $\delta$ 1 T subset, which contributes to the establishment and maintenance of early pregnancy immune tolerance through secretion of anti-inflammatory cytokines and modulation of negative signaling pathways [31]. The V $\delta$ 2 T subset becomes predominant in the decidua during mid-pregnancy, whereas V $\delta$ 1 dominance returns at term [32]. In pathological pregnancies,  $\gamma\delta$  T cells may acquire proinflammatory and cytotoxic functions, warranting further investigation [33].

Although the significance of B lymphocytes in pregnancy has not been fully clarified, existing studies suggest that they play an active role during pregnancy [34,35]. In recent years, regulatory B cells have been proposed as key participants in immune tolerance. Studies have shown that a reduction

or dysfunction of circulating regulatory B cells may be associated with adverse obstetric outcomes [36,37], as these cells are crucial in fostering the immunological milieu required for implantation and may also help re-establish fetal immune tolerance in immune-mediated pregnancy

complications. Figure 1 illustrates the dynamic changes in the maternal systemic immune microenvironment during normal pregnancy, including alterations in the quantity and function of key immune components such as neutrophils, monocytes, NK cells, T cells, and B cells.



**Fig. 1 Alterations in the maternal systemic immune system in pregnancy.**

This figure illustrates systemic immune adaptations during normal pregnancy. Peripheral T cell counts decrease compared with the pre-pregnancy state, accompanied by a dynamic Th1/Th2 shift and an increase in Tregs. Neutrophils progressively accumulate and exhibit cytokine release in association with hematopoietic factors such as G-CSF and GM-CSF, with changes between resting and activated states. Monocytes show stage-dependent functional polarization, shifting from a proinflammatory profile toward an anti-inflammatory/tolerant profile in late pregnancy. NK cells display differences between peripheral blood and the local maternal-fetal immune microenvironment. In addition, reduced or dysfunctional circulating Bregs may be associated with adverse obstetric outcomes. Th1. T helper 1; Th2. T helper 2; Tregs. Regulatory T cells; IL-17. interleukin-17; IL-6. Interleukin-6; G-CSF. Granulocyte colony-stimulating factor; GM-CSF. Granulocyte-macrophage colony-stimulating factor; TNF. Tumor necrosis factor; IL-12. Interleukin-12; NK. Natural killer; Bregs. Regulatory B cells

### Maternal-fetal immune adaptation

The endometrium (decidua) plays a key role in embryo implantation and pregnancy maintenance, with decidual immune cells being key regulators of these processes [38]. Decidual NK cells (dNK) constitute the predominant leukocyte population (50%–70%) in the decidua. Unlike peripheral blood NK cells, dNK cells exhibit minimal cytotoxicity and a functional bias toward immune regulation and angiogenesis, playing a crucial role in placental formation and fetal development [39]. During early pregnancy (8–10 weeks), dNK cells secrete angiogenic factors that promote spiral artery remodeling and optimize placental perfusion. With the progression of pregnancy, after 12–14 weeks of gestation,

dNK cells mainly secrete cytokines [e.g., interleukin (IL)-8 and IL-10], which promote extravillous trophoblast (EVT) invasion and enhance placental function by modulating matrix metalloproteinase (MMP)-9 and inhibiting EVT apoptosis [40]. Additionally, dNK cells limit excessive trophoblast invasion and protect maternal tissues by transforming growth factor (TGF)- $\beta$  and interferon (IFN)- $\gamma$  secretion, maintaining immune tolerance and placental homeostasis [41].

Macrophages also contribute significantly to the decidual immune microenvironment. While scarce in the nonpregnant endometrium, macrophage numbers rise significantly in the decidua during pregnancy, driven by chemokine-mediated recruitment. Most exhibit an alternatively activated

macrophage (M2) phenotype, expressing CD163, CD206, IL-10, IL-6, tumor necrosis factor (TNF), and chemokine C-C motif ligand (CCL)-4, a profile resembling that of macrophages stimulated by granulocyte-macrophage colony-stimulating factor and IL-10 [6]. During placentation, macrophages localize near invading trophoblasts and uterine spiral arteries and support trophoblast invasion and spiral artery remodeling by secreting and regulating MMPs, which promote extracellular matrix degradation and tissue remodeling [42]. Furthermore, decidual macrophages are crucial for maintaining immune homeostasis at the maternal-fetal interface. They not only mediate tissue clearance by phagocytosing apoptotic cells and debris but also help constrain excessive maternal immune responses through the secretion of anti-inflammatory cytokines such as IL-10 and TGF- $\beta$ . Notably, macrophages within the placental bed express inhibitory receptors that interact with HLA-G expressed on extravillous trophoblasts. The interaction between HLA-G and these macrophages reprograms their cytokine profile, fine-tuning their function to promote immune tolerance [43].

Decidual T cells, comprising 5%–20% of decidual lymphocytes in early pregnancy and increasing to 40%–80% at term, differ notably from peripheral blood T cells [44,45]. Among CD4<sup>+</sup> T cells, 10%–30% express FOXP3, indicating a significant enrichment of Tregs relative to the circulation [46]. The frequency of Th1 cells in the decidua moderately increases, whereas Th17 and Th2 cells are typically not enriched, indicating the presence of a mild inflammatory environment under the control of Tregs [47]. Decidual Tregs increase in early pregnancy, remain elevated during mid-pregnancy, and then decrease before delivery. Animal studies have demonstrated that T cells can interact with uterine NK cells, affecting the maternal hemodynamic response to pregnancy [48,49]. The negative impact of uterine NK cell deficiency on decidual vascular remodeling is exacerbated by concurrent T cell deficiency. Insufficient or dysfunctional Tregs are linked to infertility, miscarriage, preeclampsia (PE), and fetal growth restriction [50]. Notably, unlike peripheral blood where CD4<sup>+</sup> T cells predominate, at term pregnancy, CD8<sup>+</sup> T cells constitute the major T cell subset in the decidua, with most being activated effector memory T cells; their low basal secretion of perforin and granzyme B results in reduced cytolytic activity [51]. Furthermore, the robust secretion of IFN- $\gamma$  and other factors by decidual CD8<sup>+</sup> T cells suggests a positive role in pregnancy, potentially contributing to vascular processes within the decidua [52]. Maintaining a proper equilibrium between immunotolerance and antiviral responses via decidual CD8<sup>+</sup> cytotoxic T cells is essential for the normal

course of pregnancy. Figure 2 depicts the dynamic changes in the immune microenvironment at the maternal-fetal interface during normal pregnancy, highlighting dNK cells, macrophages, and T cells.

### **Autoimmune diseases during pregnancy**

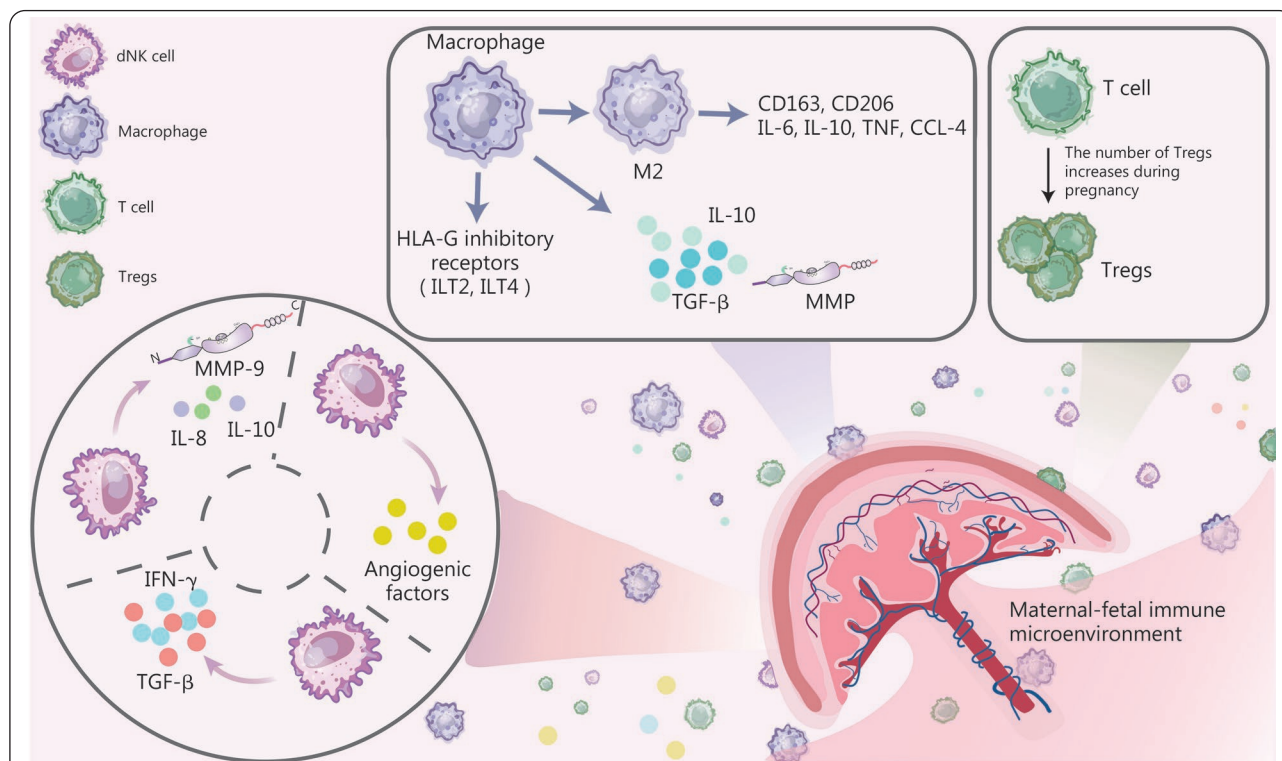
The risk of APOs is markedly elevated in pregnant women with AIDs, driven not only by disease-specific immune dysregulation [e.g., anti-double-stranded DNA (dsDNA) antibodies in systemic lupus erythematosus (SLE) and inflammatory cytokines in rheumatoid arthritis (RA)], but also by environmental factors such as hormonal fluctuations and chronic systemic inflammation [53–55]. AIDs exhibit distinct patterns of immune adaptation at both the systemic and local levels, which highlights the clinical relevance of elucidating pregnancy-related immune microenvironmental shifts in these conditions to improve patient management. Figure 3 illustrates a simplified diagram of immune alterations across common AIDs during pregnancy.

### **Systemic lupus erythematosus**

SLE is a complex, chronic autoimmune disorder characterized by widespread immune-mediated tissue injury across multiple organ systems. Clinically, it presents with manifestations such as malar rash and positive antinuclear antibody [56]. The dysregulation of both innate and adaptive immune systems constitutes a significant driving force, with aberrant B cell activation and the resulting elevated autoantibodies [including anti-dsDNA, anti-smith, and antiphospholipid (aPL) antibodies] and immune complex deposition, leading to complement activation and tissue injury [57]. In addition, T cell imbalance, which is characterized by excessive Th17 activation and impaired Treg function, further disrupts immune regulation [58].

Women with SLE have a pregnancy rate approximately 30% lower than that of the general population, and face greater maternal risks (lupus flare, PE, and thrombosis) along with fetal and neonatal morbidity [59–61]. In SLE, higher disease activity during pregnancy is a key predictor of adverse outcomes [62]. Additionally, many immunosuppressive therapies pose risks [63]. Accordingly, clinical guidelines recommend conception only after achieving stable low disease activity under treatment with low-dose steroids and/or hydroxychloroquine (HCQ) [64].

Pregnancy involves complex immunoregulatory processes, posing significant challenges for patients with SLE. In healthy pregnancies, a shift from proinflammatory Th1/Th17 dominance to anti-inflammatory Th2/Treg dominance



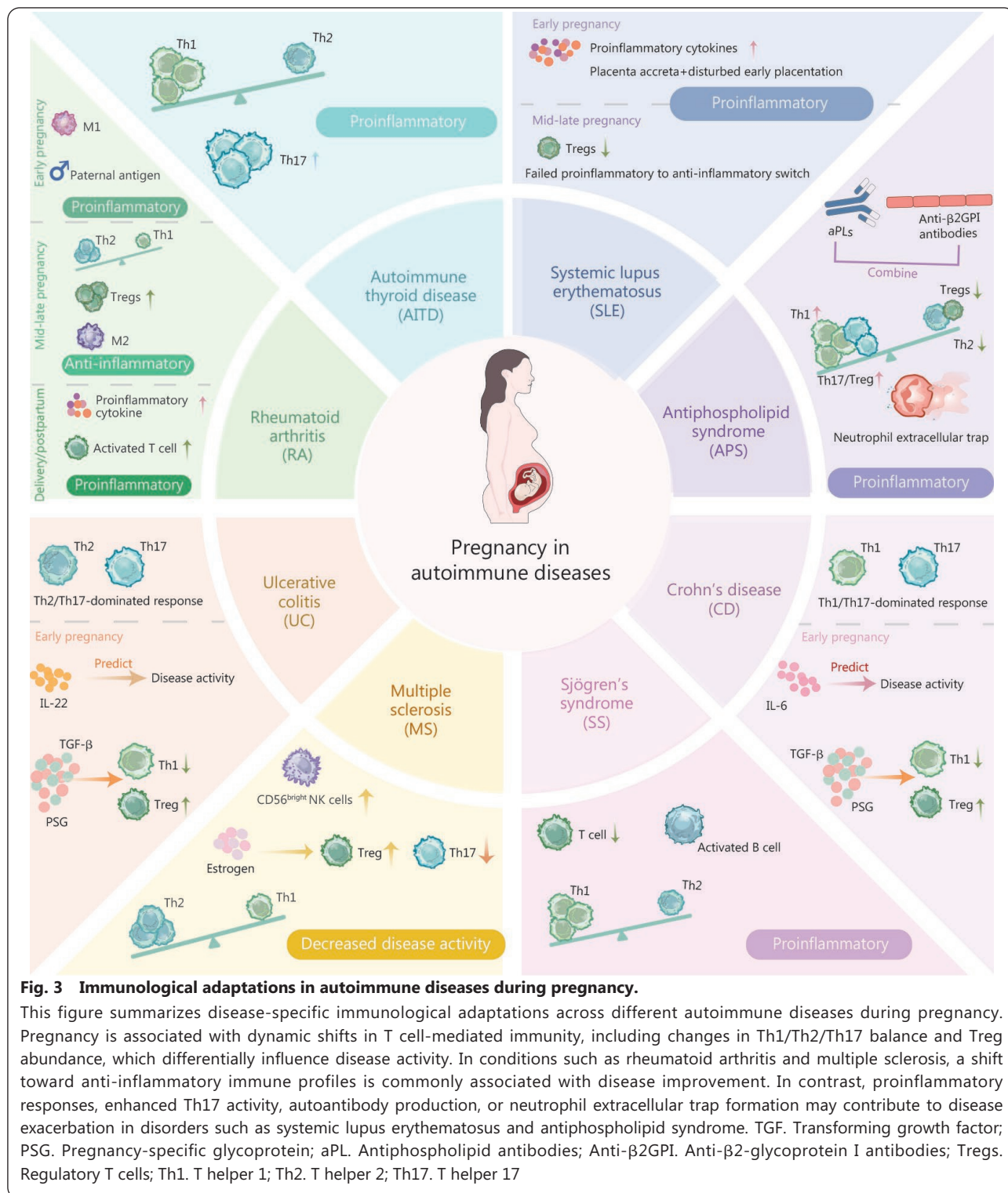
**Fig. 2 Maternal-fetal immune adjustment during normal pregnancy.**

This figure illustrates the dynamic immune microenvironment at the maternal-fetal interface during normal pregnancy. dNK cells, macrophages, and T cells constitute the major immune cell populations in the decidua. dNK cells secrete angiogenic factors, cytokines, and matrix metalloproteinases to support placental vascular remodeling. Decidual macrophages preferentially polarize toward an M2-like phenotype, expressing CD163 and CD206 and producing immunoregulatory mediators, thereby contributing to immune tolerance and tissue remodeling. Meanwhile, the proportion of Tregs increases during pregnancy, playing a central role in maintaining immune tolerance at the maternal-fetal interface. Together, these coordinated immune adaptations ensure successful implantation, placental development, and the maintenance of pregnancy. TNF. Tumor necrosis factor; TGF. Transforming growth factor; MMP. Matrix metalloproteinase; HLA. Human leukocyte antigen; CCL. Chemokine C-C motif ligand; dNK. Decidual natural killer; Tregs. Regulatory T cells; Th1. T helper 1; Th2. T helper 2; Th17. T helper 17; IL. Interleukin; ILT. Immunoglobulin-like transcript

promotes maternal-fetal tolerance. In SLE patients, this regulatory shift is disrupted due to baseline immune dysregulation, increasing pregnancy-related risks. Early pregnancy in those with SLE is characterized by elevated levels of proinflammatory cytokines that impair embryo implantation and early placental development [65]. In the middle to late stages of pregnancy, SLE patients fail to transition from a proinflammatory to an anti-inflammatory state, resulting in insufficient anti-inflammatory responses. This may lead to a sustained proinflammatory milieu within the maternal immune system, ultimately increasing the probability of complications [66]. In healthy pregnancies, mid-to-late gestation is typically features by increased anti-inflammatory cytokines and expansion of Tregs to maintain maternal-fetal immune balance, but SLE patients exhibit impaired regulation, hindering this transition [65,67]. Moreover, B cells in pregnant women with SLE show marked gene expression changes, including major histocompatibility complex pathway activation

and inflammatory factor dysregulation, which, in conjunction with aberrant interactions with T cells and monocytes, synergistically promote disease progression [65,67].

As mentioned above, the maternal immune system undergoes a transition from type 1 immunity to type 2 immunity during a healthy pregnancy, usually manifesting as an increased Th2-mediated immune response and a reduced Th1 response, thereby facilitating the sustenance of pregnancy. This immune shift may reduce disease activity in some SLE patients, resulting in “immunosuppression” rather than the typical “immune hyperactivation” phenotype. Mechanistically, a surge in estrogen facilitates Th2-mediated immune responses and inhibits Th1 responses, whereas progesterone further bolsters maternal-fetal immune tolerance by suppressing Th17 cell activity and increasing the ratio of Tregs [68]. Moreover, placental anti-inflammatory cytokines are crucial in maintaining local immune equilibrium by suppressing proinflammatory Th17 responses and enhancing



the functionality of Tregs [69]. Furthermore, changes in galactosylation may play a role in the development of SLE. Research indicates that galactosylation-related transcripts are increasingly expressed in pregnant SLE patients, with their overall expression exceeding that of healthy individuals, implying that enhanced galactosylation-related immune

characteristics may be present in SLE patients either at baseline or during gestation [70,71]. Although the molecular mechanisms underlying immune modulation during normal pregnancy have been extensively documented, studies focusing on dynamic immune regulation in pregnant SLE patients are still exceedingly scarce, and further investigations are

required to delineate the dynamic alterations in the immune microenvironment and to understand their direct effects on pregnancy outcomes.

In addition to the maternal systemic immune system, immune homeostasis within the maternal-fetal milieu is profoundly disrupted in SLE patients. Elevated aPL levels can lead to placental microangiitis and thrombosis, thereby increasing the risk of APOs [72]. Within the gestational uterus, dNK cells account for 70% of the maternal immune cell population, significantly contributing to placental development and immune equilibrium in the maternal-fetal milieu. Nevertheless, in pregnant individuals with SLE, dNK cells may exhibit functional abnormalities stemming from immune imbalance, resulting in unfavorable pregnancy outcomes [73]. However, current evidence suggests that a reduction in dNK cell numbers, enhanced cytotoxicity, or altered secretory functions may disrupt placental immune balance [74]. Nevertheless, the exact role and mechanisms of dNK cells in SLE-associated abnormal pregnancy outcomes still need to be more thoroughly investigated. Further studies should focus on the dynamic alterations and their direct links to placental dysfunction to uncover the mechanisms of local immune imbalance during pregnancy in SLE.

The application of single-cell RNA sequencing technology has offered new insights into the immune microenvironment in patients with SLE. Lien *et al.* [70] revealed that the distinct immune profile of SLE patients compared with that of healthy controls and RA patients is primarily attributed to the IFN signature characteristic of SLE. In contrast to the pregnancy-associated downregulation of IFN-related genes in healthy controls, SLE patients show marked expansion of IFN-regulated T cells (SC-T3), monocytes (SC-M2), and nonclassical monocytes (SC-M3) [70]. Notably, SC-M3 expansion is partially reversed, whereas SC-M2 gains a more prominent position in the immune cell composition [70]. These findings suggest that altered immune cell composition may contribute to disease activity and pregnancy outcomes in patients with SLE. Relevant studies indicate that SLE patients have increased disease activity at 6 and 12 months after delivery, which aligns with results from the RevNatus registry [75,76]. These results emphasize the importance of close monitoring and management during the first postpartum year, given the incomplete immune restoration. Personalized immune monitoring and treatment during pregnancy are recommended for SLE patients.

### **Rheumatoid arthritis**

SLE is one of the most common systemic AIDs encountered

during pregnancy, whereas RA, another common disease that poses different challenges in pregnancy care, is a complex autoimmune disorder characterized by immune-mediated inflammation of the synovium [77], along with cartilage and bone erosion, primarily impacting multiple joints, particularly small joints in the hands and feet. Pregnant women with RA are especially vulnerable to APOs, including higher rates of low birth weight and stillbirth [78,79]. Disease control and medication management critically influence pregnancy outcomes [80-82].

Immune cells, especially CD4<sup>+</sup> T cells (e.g., Th17 cells), B cells, synovial macrophages, and synovial fibroblasts, are highly accumulated in the local synovium, and participate in chronic inflammation and bone destruction [83,84]. T cells in RA patients exhibit abnormal activation and differentiation. For example, substantial quantities of proinflammatory cytokines are generated by Th17 cells and CD4<sup>+</sup>CD28<sup>-</sup> T cells, thereby exacerbating joint inflammation [85]. Furthermore, autoreactive B cells contribute by producing rheumatoid factor and anti-citrullinated protein antibodies (ACPAs), forming immune complexes that deposit in joints, activate complement, and exacerbate inflammation and tissue damage [86]. These alterations in the immune microenvironment collectively promote persistent inflammation and joint damage in RA.

In contrast to SLE, RA displays unique immune changes and a distinct disease course throughout pregnancy. This phenomenon of symptom remission, first observed by Hench in 1938, has since been confirmed by multiple studies [87-89]. Extensive research indicates that 75%–95% of RA patients undergo remission of disease activity, which is most evident in early pregnancy, peaks in late pregnancy [90,91], whereas approximately half of patients experience disease flares postpartum [92]. Multiple mechanisms within the immune system help to contribute to the pregnancy-related improvement of RA, whereas the cessation of postpartum immunoregulatory processes is associated with a flare-up of the disease after delivery. Maternal-fetal HLA disparities, immunoglobulin (Ig)G galactosylation, and immunoregulatory mechanisms, along with alterations in innate and adaptive immune cells and their associated cytokines, are pivotal factors in improving RA symptomatology during pregnancy [92-94]. IgG galactosylation is a pregnancy-associated phenomenon that reduces the pathogenicity of disease-specific autoantibodies. An increase in the galactosylation of anti-cyclic citrullinated peptide IgG during pregnancy is linked to the improvement of RA [95]. In addition, research has shown that patients who test positive for ACPAs experience more limited improvement in RA disease activity [92,96].

ACPAs represent the most RA-specific autoantibody, and alterations in galactosylation pattern are strongly tied to disease activity [95,97]. Bondt *et al.* [98] discovered that ACPA-IgG galactosylation was markedly elevated, correlating with enhanced Disease Activity Score in 28 joints using C-reactive protein (DAS28-CRP) levels in RA patients positive for the autoantibody. Meanwhile, galactosylation changes in total IgG did not significantly correlate with disease improvement, suggesting that galactosylation of disease-specific autoantibodies such as ACPA-IgG, is pivotal in the alleviation of RA symptoms [98]. Indeed, galactosylation changes are implicated in the pathogenesis of AIDs. In addition to RA, disorders such as inflammatory bowel disease (IBD), multiple sclerosis (MS), and autoimmune thyroid disease (AITD) also exhibit galactosylation abnormalities [99,100]. Nevertheless, alterations in galactosylation in these diseases have yet to be reported.

This immune trajectory helps explain why RA often improves during pregnancy but flares after delivery. During the initial phase of pregnancy, paternal antigens (e.g., sperm, TGF- $\beta$ , prostaglandins, soluble HLA molecules, and other bioactive molecules) and M1 macrophages drive a proinflammatory state to promote implantation [101,102]. To prevent fetal rejection, FOXP3<sup>+</sup> Tregs and M2 macrophages are subsequently induced, a Th2-type immune response is adopted, effector T cells are downregulated, and anti-inflammatory cytokines (e.g., IL-10) are produced, leading to an immune tolerance state during pregnancy. As gestation approaches delivery, the immune balance shifts back to a proinflammatory state in preparation for delivery. Within 12 weeks postpartum, CD4<sup>+</sup> and CD8<sup>+</sup> T cells exhibit increased activation, along with increased levels of inflammatory mediators. Prolactin activates B cells, promotes the secretion of proinflammatory cytokines, modulates Treg function, and stimulates macrophages to release multiple cytokines [103,104]. Additionally, recent findings highlight that microchimeric cells may initiate autoimmune responses by presenting as persistent cells or antigens in the maternal system, acting as sources of RA-specific autoantibodies and participating in the activation or aggravation of postpartum autoimmune reactions [105]. This mechanism provides new insights into understanding pregnancy-associated variations in RA disease progression.

RNA sequencing and gene expression analyses provide insights into immune microenvironment changes in pregnant RA patients, particularly regarding IFN-I signaling, B cell function, and neutrophil activity. Studies have shown that women with RA who experience symptom improvement

exhibit a marked upregulation of IFN-I-induced genes, whereas those whose symptoms worsen do not show such changes [106-108]. These findings highlight the role of IFN-I signaling in modulating immune tolerance and inflammatory responses in pregnancy-related RA progression. RNA-seq analysis revealed distinct pre-pregnancy (T0) gene expression profiles between the improvement and deterioration groups. Co-expression network analysis showed that B cell-associated genes, including CD19, CD22, CD79A/B, were enriched in the deterioration group, suggesting that B cells drive adverse immune responses [109]. Conversely, neutrophil-related genes, including folate receptor gamma (FOLR3), alanyl aminopeptidase (ANPEP), were upregulated in the improvement group, indicating enhanced neutrophil activity [70]. Collectively, these findings emphasize dynamic immunological shifts during pregnancy and the critical roles of B cells and neutrophils in RA pathogenesis.

Despite existing studies providing robust evidence, gaps remain concerning the changes in the local immune microenvironment of RA patients, particularly in the placenta and decidua. Unlike other AIDs, RA often shows a characteristic course of improvement during pregnancy followed by postpartum relapse. This distinctive “pregnancy remission-postpartum relapse” phenomenon has attracted extensive research attention [110,111]. However, most studies have focused on systemic immune changes, while the specific role of the local immune microenvironment in RA remains unclear. Clarifying these mechanisms will enable the development of more targeted immunoregulatory strategies for RA management.

### **Antiphospholipid syndrome**

Unlike the systemic inflammation driven by joint involvement in RA, the pregnancy risks associated with antiphospholipid syndrome (APS) are mainly attributable to aPLs and blood hypercoagulability, rather than direct inflammatory damage. APS is an autoimmune disorder predominantly characterized by recurrent thrombosis and APOs. Pregnant women with APS face greater risks for both maternal and fetal outcomes [112,113]. Maternal complications commonly include pregnancy-induced hypertension, PE, and thromboembolic events [114-116]. Fetal outcomes include significantly higher incidences of miscarriage, preterm birth, intrauterine growth restriction (IUGR), and stillbirth [117]. Research has indicated that the risk of APOs in APS patients can reach approximately 50% [118], and these risks are closely related to the types and titers of aPLs as well as other clinical manifestations (SLE) [117,119]. The key pathological marker

of APS is the presence of aPLs. By binding to phospholipids, aPLs form immune complexes that trigger complement activation, platelet aggregation, and endothelial injury, thus increasing the risk of thrombosis. In addition, aPLs further stimulate inflammatory responses and immune complex formation by activating the complement system, ultimately causing endothelial damage and thrombosis [120,121]. While aPLs are characteristic of APS, they can also be detected in other AIDs, such as SLE [122], implying that these antibodies play an important part in the immune responses of those ailments as well.

aPLs significantly influence the placenta and the maternal immune system in APS patients. The high expression of  $\beta$ 2-glycoprotein I ( $\beta$ 2GPI) in the placenta makes it a primary target for aPLs, and its binding can induce endothelial cell phenotypic changes, platelet aggregation, and coagulation dysfunction, with complement system activation acting as a critical mediator in this process [123,124]. This pathological change reduces trophoblast invasion capacity, resulting in early pregnancy loss; if pregnancy continues, a poorly perfused placenta may lead to complications such as PE and IUGR [125].

As key effector cells of innate immunity, neutrophils exhibit abnormal activation in pregnant women with APS, leading to a significant increase in the release of neutrophil extracellular traps (NETs). These NETs participate in autoimmune pathogenesis and directly impair normal placental development by suppressing trophoblast invasion and migration capabilities [126]. Furthermore, NETs may negatively influence the migratory and tubulogenesis capacities of human umbilical vein endothelial cells, further increasing the likelihood of thrombosis. Moreover, during the adaptive immune response,  $\beta$ 2GPI-reactive  $CD4^+$  T cells accelerate immune dysregulation by activating B cells, leading to enhanced secretion of anti- $\beta$ 2GPI antibodies. aPLs can also bind to their target antigen  $\beta$ 2GPI, activating membrane receptors and downstream signaling pathways, which can in turn lead to NK cell activation [127]. In both primary antiphospholipid syndrome (PAPS) and secondary antiphospholipid syndrome (SAPS), research has shown that patients present significantly elevated absolute counts of total T cells and  $CD4^+$  T cells in peripheral blood, along with a marked Th1/Th2 imbalance manifested by increased Th1 cells, reduced Th2 cells, decreased Tregs, and an increased Th17/Treg ratio [128]. Notably, alterations in T cell subsets in APS patients are strongly correlated with their clinical phenotypes. Patients with thrombotic APS often present elevated proportions of activated  $CD4^+HLA-DR^+$  and  $CD8^+HLA-DR^+$  T cells, characterized by the upregulation of HLA-DR [129]. Among obstetric APS patients, those without

thrombosis present higher proportions of  $CD4^+CD45RA^+CCR7^+$  memory T cells and a higher frequency of activated  $CD4^+CD25^+$  subsets [130]. These findings suggest that T cell changes in thrombotic APS are more closely linked to proinflammatory and pro-thrombotic processes, whereas obstetric APS may involve specific regulation of the placental immune microenvironment.

B cells are key contributors to the development of APS. Research has shown that peripheral naive B cells are significantly elevated in both proportion and absolute number, whereas memory B cells are reduced. This abnormal pattern correlates with low complement C4 levels and high titers of IgG anticardiolipin and anti- $\beta$ 2GPI antibodies [130]. Naive B cells potentially facilitate disease progression via antigen presentation and autoantibody production. Their increased numbers may reflect defective B cell trafficking, peripheral overactivation, or abnormal differentiation into plasma cells, causing memory B cell loss. These findings indicate that B cell subset imbalance is a critical immunologic hallmark of thrombosis in APS patients and a potential factor contributing to poor pregnancy outcomes in affected patients.

aPLs contribute to pregnancy complications through multiple pathways, and their pathogenic effects extend beyond thrombosis. For example, anti- $\beta$ 2GPI antibodies bind to  $\beta$ 2GPI on endothelial cells, triggering the coagulation cascade, inducing tissue factor expression, and promoting platelet activation and aggregation, thereby driving microthrombi formation in placental and umbilical vessels, impairing placental perfusion, and ultimately increasing the risk of adverse pregnancy outcomes [131]. Moreover, aPLs can specifically disrupt the anticoagulant annexin V barrier on trophoblast surfaces [132,133], increasing the susceptibility of placental cells to abnormal apoptosis and differentiation disorders [134]. Concurrently, aPLs induce local inflammation via complement activation and promotes proinflammatory cytokine secretion, contributing to a prothrombotic and proinflammatory microenvironment. Nevertheless, the precise molecular mechanisms through which aPLs contribute to pregnancy complications have yet to be fully elucidated. Further studies are needed to clarify these unresolved mechanisms, which will provide theoretical support for the development of novel targeted therapies to inhibit the pathogenicity of aPLs.

### **Sjögren's syndrome**

Sjögren's syndrome (SS) is a chronic immune-mediated disease that predominantly targets the salivary and lacrimal glands, resulting in prominent dryness of the mouth, eyes, and skin, and is often accompanied by fatigue, joint pain, and

neuropsychiatric symptoms. Investigations into reproductive outcomes in primary Sjögren's syndrome (pSS) patients have yielded mixed and somewhat controversial findings. A multicenter, prospective cohort study in France indicated that, relative to people in general, women with pSS generally have a better prognosis during spontaneous pregnancy, with no substantial elevation in the overall risk of APOs [135]. However, for pSS patients undergoing *in vitro* fertilization, a multicenter retrospective cohort study found that they had poorer fertility and clinical pregnancy status, especially those characterized by oocyte and embryo developmental abnormalities [136]. This discrepancy suggests that pSS may impact pregnancy outcomes differently under natural conception and assisted reproductive technology, indicating potential mechanistic differences that warrant further investigation. Beyond maternal pregnancy outcomes, pSS also carries fetal and neonatal risks mediated by autoantibodies. Mothers carrying anti-Sjögren's syndrome-related antigen A (SSA/Ro) and anti-Sjögren's syndrome-related antigen B (SSB/La) autoantibodies are at increased risk of neonatal lupus-related cardiac complications, with congenital heart block (CHB) being the most severe complication [137].

The main characteristic of SS is immune system dysregulation, particularly with respect to aberrant T and B cell activation. In SS patients, T cell infiltration, with a predominance of CD4<sup>+</sup> subsets, is markedly evident within the salivary glands [138,139]. Research indicates that multiple T cell subtypes, including Th1, Th2, Th17, and Tregs, are fundamentally involved in the onset and progression of SS [140,141]. Th1 cell activation enhances the secretion of inflammatory mediators. Moreover, Th2 cells spur excessive B-cell activation by releasing cytokines, which in turn boost the generation of autoantibodies (e.g., anti-SSA/Ro and anti-SSB/La), exacerbating the immune response in SS [142,143].

Current research on the changes in the immune microenvironment of SS during pregnancy is still quite limited. Autoantibodies, especially anti-SSA and anti-SSB, are present in most SS patients and can traverse the placental barrier, potentially triggering immune complications in the fetus. Upon fetal exposure to maternal Ro/La antibodies, these antibodies can be actively transferred to the fetal circulation via trophoblastic Fc $\gamma$  receptors. These antibodies recognize SSA/Ro-SSB/La antigens exposed on fetal cardiomyocytes due to physiological apoptosis during cardiac development, triggering proinflammatory phagocytic responses that lead to irreversible fibrotic injury of the conduction system [144]. The risk of neonatal lupus-related cardiac complications is elevated, with CHB representing the most severe complication [145].

Ivanchenko *et al.* [146] noted changes in B-cell subpopulations, relatively fewer T cells, and lower NK cell frequencies in Ro/La<sup>+</sup> mothers, which aligns with the immunological features observed in SS. Notably, in newborns exposed to Ro/La antibodies, the frequency of CD56<sup>dim</sup>CD16<sup>high</sup> NK cells was increased, suggesting that their NK cells may be activated and exhibit cytotoxic activity. IFN- $\alpha$  is considered critical to this pathway because it drives NK cell proliferation via antibody-dependent cellular cytotoxicity (ADCC), NK cells bind to Ro/La antibodies through CD16, resulting in myocardial cell injury and culminating in CHB.

The interactions between Ro/La antibodies, IFN, and cytotoxic cells may also constitute one of the core mechanisms underlying the pathogenesis of this disease. These immune mechanisms not only influence the maternal immune microenvironment but also affect the immune response of newborns through the placenta [145]. Thus, investigating the function of Ro/La antibodies reveals the intricate alterations in immune cell groups during pregnancy, offering new insights into the mechanisms underlying SS-related pregnancy complications and potentially providing theoretical support for future diagnostic and treatment strategies.

### Multiple sclerosis

MS, a chronic AID of the central nervous system, presents a distinct pregnancy-associated immune profile. MS is characterized by immune-mediated demyelination and axonal injury, driven by CD4<sup>+</sup> T cells (Th1, Th17), glial cells, B cells, dendritic cells, and macrophages [147-149]. Th1 and Th17 cells promote central nervous system inflammation through cytokine secretion, whereas B cells exacerbate demyelination via antibody production and complement activation [150-152]. Dendritic cells and macrophages enhance immune cell activation and migration, and glial cell dysfunction under immune dysregulation may contribute to neuronal injury [153,154]. This dysregulated immune microenvironment sustains neural damage and disease progression.

During pregnancy, the immune microenvironment of MS experiences intricate and important alterations, significantly affecting disease activity and clinical presentation. Hormonal fluctuations, particularly in estrogen, progesterone, and testosterone, contribute to a reduced relapse rate [155,156]. Estradiol limits encephalitogenic T cell activation and central nervous system infiltration via its  $\alpha$  receptor, exerting neuroprotective effects, whereas estriol enhances Treg expansion and suppresses Th17 differentiation, further alleviating disease activity [157,158]. However, these immunomodulatory effects are often disrupted after delivery,

and many MS patients experience a transient worsening of their condition postpartum [159]. This phenomenon suggests that the immune tolerance and regulatory effects observed are transient and that postpartum immune tolerance may revert to a heightened inflammatory state. In addition to hormonal influences, IFN may also regulate disease activity in MS patients. While IFN-I signatures are linked to disease activity in SLE, IFN biology in MS appears more nuanced: exogenous IFN-I has well-established therapeutic efficacy in MS, whereas the contribution of endogenous IFN signaling to pregnancy-related remission and postpartum relapse remains unclear. Notably, an increased IL-10/IFN- $\gamma$  ratio in late pregnancy may promote immune tolerance, potentially representing another mechanism underlying MS remission [160]. Furthermore, a similar “pregnancy remission-postpartum relapse” pattern observed in RA patients suggests that pregnancy-related immune regulation may involve shared pathways across different AIDs [108,161]. A comprehensive exploration of the involvement of IFN in pregnancy-associated AIDs could advance the understanding of MS remission mechanisms and potentially offer new therapeutic targets for preventing postpartum relapse.

In patients with MS, autoreactive T cells activated in the periphery demonstrate significant abnormalities, most notably those of the CD4<sup>+</sup> subset [162,163]. Among untreated, nonpregnant individuals with MS, the peripheral blood shows a considerably increased level of CD4<sup>+</sup> T cells compared with that of healthy controls [164], which is in line with the characterization of MS as a CD4<sup>+</sup> T cell-mediated immunopathology. Notably, during pregnancy, individuals with MS continue to display an elevated proportion of CD4<sup>+</sup> T cells, with no significant changes observed compared to the nonpregnant period. This may obscure the alterations in immune cell subsets involved in immune regulation [164]. For instance, MS patients exhibit a shift from Th1-type to Th2-type immune responses, which reverts back to Th1-type responses after delivery [160,165]. This shift may be a contributing factor to the exacerbation of MS symptoms postpartum. In addition to Th cells, the function of Tregs in MS remains controversial [166-168]. Many investigations have suggested that in healthy women, Treg counts drop during gestation and return to pre-pregnancy levels postpartum. However, in women with MS, the Treg frequency remains constant throughout pregnancy, without a significant postpartum rebound or decline relative to the the Treg frequency in nonpregnant women [169]. These observations conflict with some reports indicating that Treg counts in pregnant MS patients may either increase or decrease [170]. Importantly, many studies rely on FOXP3 as a Treg

marker, and despite FOXP3 being critical for Treg development and function, it can also be upregulated in newly activated conventional T cells, leading to potential misinterpretation when identifying and characterizing Tregs [171-173].

NK cells also contribute to immune modulation in pregnant MS patients. The peripheral NK cell profiles shift, with increased CD56<sup>bright</sup> and decreased CD56<sup>dim</sup> subsets, mirroring the changes observed in healthy pregnancies [174]. These shifts typically reverse postpartum, indicating that pregnancy-induced NK cell regulation is transient. However, in MS patients, CD56<sup>bright</sup> NK cells have a reduced capacity to suppress autologous CD4<sup>+</sup> T cell proliferation compared with that in healthy controls [169], suggesting impaired regulatory function, which may limit clinical improvement during pregnancy.

Locally, similar changes occur in the decidua. CD56<sup>bright</sup> NK cells, although enriched in the decidua relative to those in peripheral blood, are lower in pregnant MS patients than in healthy pregnant women, implying compromised immunoregulation [169]. Decidual macrophages predominantly adopt an M2 phenotype, derived from CD14<sup>+</sup>CD163<sup>+</sup> monocytes, contributing to immune tolerance, and resembling anti-inflammatory macrophages found in MS [175,176]. In addition, placental extracellular vesicles and decidual mesenchymal stem cells play vital roles in immunoregulation. Extracellular vesicles maintain immune tolerance by modulating the proliferation and differentiation of immune cells, whereas decidual mesenchymal stem cells inhibit Th1 and Th17 cells and enhance Treg activity, further supporting immune tolerance during gestation [177,178].

In recent years, emerging technologies have offered pivotal insights into the immune microenvironment during pregnancy in MS. Epigenomic and transcriptomic studies have shown that T cells in MS patients display distinctive epigenetic and transcriptional alterations, correlating with disease remission and reversal after delivery [179,180]. Moreover, proteomic analysis further revealed a characteristic profile in which immunomodulatory proteins, such as programmed death ligand 1 (PD-L1) and leukemia inhibitory factor receptor (LIF-R), were upregulated, whereas inflammatory mediators, including C-C motif chemokine ligand 8 (CCL8) and C-X-C motif chemokine ligand 5 (CXCL5), were downregulated in gestation [181], offering critical insights for understanding pregnancy-related immune regulation and identifying novel biomarkers.

### Other autoimmune diseases

The immunomodulatory impact of pregnancy is not limited

to prototypical autoimmune rheumatic diseases, it can also be observed in other autoimmune conditions, such as IBD and AITD. Despite having different pathogenic mechanisms, IBD and AITD both exhibit unique immune regulation patterns during pregnancy.

IBD is a chronic disorder characterized by intestinal inflammation, including Crohn's disease (CD) and ulcerative colitis (UC). CD is mainly characterized by transmural inflammation mediated by Th1/Th17 immune responses, whereas UC predominantly involves a Th2/Th17 mixed immune response, with lesions typically confined to the colonic mucosa [182]. The risk of disease activity also differs between IBD subtypes, reaching up to 70% in UC compared with 54% in CD, although this difference is not statistically significant [183]. Biomarker profiles also reflect subtype-specific patterns. IL-6 levels in early pregnancy may predict CD activity, whereas IL-22 levels correlate with UC progression [184]. Moreover, sex hormones and immunoregulatory factors significantly influence the immune microenvironment in patients with IBD. In Th1-type experimental colitis, exogenous estrogen can reduce inflammation by downregulating proinflammatory cytokines and mast cell proteases via an estrogen receptor (ER)-dependent mechanism [185]. Placental trophoblast-derived pregnancy-specific glycoproteins (PSGs) and TGF- $\beta$  contribute to anti-inflammatory regulation by downregulating Th1-mediated inflammation and enhancing Treg populations. Nonetheless, active IBD is significantly linked to poor pregnancy outcomes [186], indicating that physiological immune adaptations may inadequately counterbalance the pathological processes of IBD.

AITD displays unique immunoregulatory features during pregnancy, mainly encompassing two prevalent subtypes, Hashimoto's thyroiditis (HT) and Graves' disease. The immune system of AITD patients undergoes changes that may impact maternal health, pregnancy outcomes, and fetal development. AITD is closely associated with APOs, including placental abruption, spontaneous miscarriage, preterm birth, pregnancy-induced hypertension, and IUGR [187]. Notably, elevated levels of thyroid peroxidase antibodies and thyroglobulin antibodies are strongly linked to a greater risk of gestational hypertension and are negatively correlated with neonatal birth weight [188]. Therefore, the immune microenvironment of AITD patients not only affects maternal disease activity but also influences pregnancy outcomes and fetal development through various mechanisms. Mechanistically, AITD is characterized by coordinated dysregulation of innate and adaptive immunity. Dendritic cells and macrophages initiate T cell activation through the

presentation of thyroid antigens, orchestrating the immune responses of Th1, Th17, and Treg subsets. Macrophage infiltration within the thyroid can induce localized chronic inflammation in HT patients, thereby intensifying thyroid dysfunction [189]. Furthermore, macrophages are capable of producing proinflammatory cytokines, which amplify inflammatory reactions and further facilitate the recruitment and activation of immune cells, thus promoting immune-mediated injury [189,190]. In patients with Graves' disease, neutrophils exacerbate thyroid damage and inflammation by releasing NETs and secreting proinflammatory factors [191]. Although the immune system generally shifts towards an anti-inflammatory state, Th1 and Th17 activation may persist in patients with AITD. This persistence may be related to fluctuations in pregnancy-associated hormones and contribute to disease relapse or worsening during pregnancy and the postpartum period [192]. Moreover, the function of NK cells may also change. Although studies indicate that their numbers may decrease, this does not imply a loss of their role in immune regulation. Various subsets of NK cells exert immunoregulatory functions, potentially having a significant impact on immune microenvironment regulation in AITD [193,194].

### **Clinical management and treatment strategies**

The clinical management of AIDs during pregnancy should consider both maternal health and fetal safety. Personalized treatment approaches are essential for effectively managing the disease while minimizing the potential adverse effects of medications on the fetus. The primary goal is to achieve a balance between controlling the disease and ensuring the safety of both the mother and the fetus. Consequently, pre-pregnancy assessments, continuous monitoring and diligent follow-up are essential elements of the management plan. Table 1 summarizes the clinical management strategies for common AIDs during pregnancy, including preconception evaluation and pregnancy management [64,184,185,195-200].

### **Pre-pregnancy evaluation and preparation**

A comprehensive pre-pregnancy assessment is crucial to ensure maternal and fetal safety. For patients with AIDs complicated by pregnancy, a joint evaluation by rheumatology and obstetrics specialists should be conducted before conception to ensure that the disease is in a stable or remission phase. Studies have shown that after receiving counseling and treatment before pregnancy, the majority of patients achieve disease remission in late pregnancy, and pregnancy outcomes are significantly improved compared with those of pregnant women who do not receive preconception counseling and

**Table 1 Clinical management strategies for common autoimmune diseases (AIDs) during pregnancy**

Disease type	Pre-pregnancy assessment and preparation	Monitoring and follow-up during pregnancy	Precautions during pregnancy	References
SLE	<p>Conduct pre-pregnancy risk stratification to ensure the disease has been in remission for ≥6 months, and adjust medications as needed (for example, discontinue cyclophosphamide);</p> <p>All SLE patients should continue taking HCQ during pregnancy;</p> <p>For individuals with no plan to become pregnant, provide contraceptive information</p>	<p>Conduct regular multidisciplinary follow-up to evaluate lupus disease activity and potential complications;</p> <p>For pregnant women who test positive for anti-Ro/SSA and/or anti-La/SSB antibodies but have no history of infants with CHB or NLE, it is recommended to perform weekly fetal echocardiography starting from 16–18 weeks until 26 weeks</p>	<p>SLE patients during pregnancy should be closely monitored for both fetal and maternal well-being, and the use of medications harmful to the fetus should be avoided</p>	[184,185,195]
RA	<p>Perform pre-pregnancy antibody screening (anti-Ro/SSA, anti-La/SSB, and aPL antibodies);</p> <p>Evaluate pre-pregnancy factors such as disease activity, use of teratogenic medications, and obstetric history;</p> <p>Discuss pregnancy plans before conception and strive to achieve clinical remission through pharmacological control of the disease. It is recommended that disease remain stable for at least six months prior to pregnancy;</p> <p>Provide contraceptive information to those who are not planning to conceive</p>	<p>Monitor joint function and regularly assess disease activity</p> <p>Evaluate medication safety and avoid drugs harmful to the fetus;</p> <p>Monitor weight gain and blood pressure levels throughout pregnancy;</p> <p>For pregnant women positive for anti-Ro/SSA and/or anti-La/SSB antibodies but without a history of infants with CHB or NLE, weekly fetal echocardiography is recommended starting at weeks 16–18 and continuing through week 26</p>	<p>Avoid medications that may affect fetal development, such as biologics, prior to delivery. During pregnancy, give priority to disease control and fetal safety</p>	[64]
APS	<p>Assess early risk factors before pregnancy (such as AIDs, history of miscarriage, thrombosis, aPL antibodies, and hypocomplementemia);</p> <p>Discuss pregnancy plans before conception and aim to achieve clinical remission of the disease through pharmacological management. It is recommended to maintain stable disease activity for at least six months prior to pregnancy;</p> <p>For individuals not planning to conceive, provide information on contraception</p>	<p>From pregnancy through 6–12 weeks postpartum, prophylactic doses of LMWH and low-dose NSAIDs should be administered; for vascular APS patients, therapeutic doses of heparin plus low-dose ASA are indicated during pregnancy.</p> <p>Women positive for aPL antibodies but without prior APS history should receive low-dose ASA to reduce the risk of preeclampsia;</p> <p>During anticoagulation, monitor coagulation parameters, platelet counts, and liver/kidney function;</p> <p>In the mid to late stages of pregnancy, monitor for diseases associated with poor placental function;</p> <p>For pregnancies involving APS, it is essential to monitor maternal and fetal health continuously and decide on the best timing for delivery</p>	<p>Pregnant APS patients face elevated risks of placental dysfunction and miscarriage, necessitating close surveillance for thrombosis and fetal well-being</p>	[199]
pSS	<p>Discuss pregnancy plans before conception and aim to achieve clinical remission of the disease through pharmacological management. It is recommended to maintain stable disease activity for at least six months before pregnancy;</p> <p>Perform a preconception assessment including factors like aPL antibodies, anti-SSA/Ro antibody profiles, disease activity, use of teratogenic drugs, history of miscarriages, and thrombosis;</p> <p>Recommend that pregnant women adjust to pregnancy-compatible medications and supplement specific vitamins</p>	<p>All pregnant women positive for anti-SSA/Ro antibodies should take HCQ;</p> <p>For pregnant women who test positive for anti-Ro/SSA and/or anti-La/SSB antibodies but have no history of infants with CHB or NLE, it is recommended to perform weekly fetal echocardiography starting from weeks 16–18 until week 26;</p> <p>Perform routine prenatal check-ups, which should include monitoring blood pressure, assessing fetal growth, and evaluating cardiac function</p>	<p>For individuals testing positive for anti-Ro/SSA and anti-La/SSB antibodies, conduct fetal electrocardiogram monitoring during early pregnancy, with management jointly handled by obstetricians and pediatric cardiology experts;</p> <p>Regularly monitor blood pressure to promptly detect and manage hypertension-related complications;</p> <p>Promote and support breastfeeding</p>	[200]

(Continued)

Disease type	Pre-pregnancy assessment and preparation	Monitoring and follow-up during pregnancy	Precautions during pregnancy	References
MS	Females diagnosed with MS should give precedence to annual preconception consultations, particularly if they are undergoing treatment or intend to begin therapy. Prior to pregnancy, engage in discussions about conception plans and endeavor to manage the disease with medications to achieve clinical remission. It is advised to ensure disease stability for a minimum of 6 months before attempting pregnancy. For patients with high disease activity, it is generally recommended to postpone pregnancy. Perform an extensive preconception assessment of the patient's general health to confirm the absence of significant complications and ensure that folic acid supplementation is initiated prior to pregnancy, thereby minimizing the risk of neural tube defects in the fetus	During pregnancy, closely track any changes in patients' symptoms. In the event of an acute flare-up, early assessment and intervention are essential. Avoid using medications that may pose potential risks to the fetus during pregnancy. If treatment is necessary, consider using medications that are relatively safer for use during pregnancy. Pregnant MS patients might face an increased number of obstetric complications, requiring diligent monitoring and timely interventions from obstetric care providers	Pregnant patients with MS require frequent follow-ups, including disease monitoring and fetal surveillance. Postpartum depression poses a potential risk for individuals with MS, underscoring the importance of psychological evaluations and supportive therapies. It is recommended that pregnant individuals with MS engage in breastfeeding after childbirth and suitably modify disease-modifying treatments and other therapeutic interventions	[196-198]

SLE. Systemic lupus erythematosus; HCQ. Hydroxychloroquine; CHB. Congenital heart block; NLE. Neonatal lupus erythematosus; RA. Rheumatoid arthritis; SSA/Ro. Sjögren's syndrome-related antigen A; SSB/La. Sjögren's syndrome-related antigen B; aPL. Antiphospholipid; APS. Antiphospholipid syndrome; AID. Autoimmune disease; LMWH. Low-molecular-weight heparin; NSAID. Non-steroidal anti-inflammatory drug; ASA. Aminosalicylic acid; pSS. Primary Sjögren's syndrome; MS. Multiple sclerosis

disease treatment [201-203]. According to the latest guidelines from the European League Against Rheumatism (EULAR), it is necessary to confirm that patients exhibit no active organ involvement before pregnancy, modify medications that are teratogenic, and ensure a safe pregnancy only when the disease is well-controlled [204]. For example, the American College of Rheumatology (ACR) advises that all women diagnosed with SLE should take HCQ during pregnancy whenever feasible [204]. Research indicates that the combination of HCQ and low-dose aspirin significantly enhances pregnancy outcomes in SLE patients by regulating the Th1/Th2 cytokine equilibrium [205]. If the patient is already taking HCQ, it is recommended to continue its use; if not, HCQ should be initiated, provided that no contraindications are evident. Moreover, the Global Antiphospholipid Syndrome Score (GAPSS) functions as a clinical tool capable of accurately predicting the response of aPL antibody-positive patients to combined aspirin and low-molecular-weight heparin (LMWH) therapy [206]. Additionally, performing both medical and investigative assessments to gauge overall disease progression and maternal-fetal risks, such as assessing disease activity (e.g., complement levels), screening for anti-Ro/SSA and anti-La/SSB antibodies, and testing for aPL antibodies, is vital. These assessments facilitate the creation of personalized treatment regimens and the adjustment of pregnancy management strategies as needed. For patients deemed high-risk, it is advisable to postpone pregnancy until the disease is sufficiently managed, usually necessitating a minimum of 6 months of pregnancy management strategies to mitigate potential risks during gestation.

### Prenatal monitoring and follow-up

Throughout pregnancy, rigorous monitoring is needed to detect and manage shifts in disease activity and fetal development in a timely manner. Periodic clinical assessments, alongside lab work (e.g., hepatic/renal function and antibody titers), can aid in gauging disease control. For example, because active disease can affect both maternal health and pregnancy outcomes, the ACR strongly recommends at least one assessment of SLE disease activity during pregnancy, including a review of clinical history, physical examination, and laboratory testing [199].

For pregnant women with anti-Ro/SSA and/or anti-La/SSB antibodies but without a history of infants with CHB or neonatal lupus erythematosus (NLE), it is recommended to perform serial fetal echocardiography from 16 to 18 weeks, continuing through 26 weeks [199]. For women with a history of having CHB or other NLE infants, weekly

fetal echocardiography should be performed from 16 to 26 weeks to facilitate early detection of potential fetal cardiac abnormalities and ensure timely intervention [199,204]. This form of surveillance helps detect possible pregnancy-related complications early and secures optimal management strategies throughout pregnancy. Notably, recent studies have confirmed that elevated peripheral blood IFN- $\alpha$  protein levels in pregnant SLE patients are independently associated with low birth weight in neonates [207,208]. However, the prognostic utility of IFN signaling for routine risk stratification requires confirmation in large-scale prospective multicenter studies.

### Special case management

Throughout pregnancy, certain patients with autoimmune rheumatic disorders may undergo notable fluctuations in disease activity or develop pregnancy-associated complications, requiring prompt and effective treatment adjustments to safeguard both the mother and fetus. For example, pregnant RA patients can experience variations in disease status, presenting with either worsening of, or alleviation of, their joint inflammation. To reduce risks to the fetus, individuals with RA are advised to cease methotrexate (MTX) prior to conception and use medications deemed safe for fetal health during pregnancy, such as sulfasalazine or HCQ, with dosage modifications made to sustain disease control [64]. When RA patients develop severe pregnancy complications, such as acute arthritis or organ involvement, a multidisciplinary team composed of rheumatologists, obstetricians, and other relevant specialists should collaborate to devise an individualized treatment plan, potentially including the use of fetal-safe corticosteroids and biologics [64]. Moreover, high-risk pregnancy conditions, including PE and placental abruption, require careful monitoring by obstetricians, and urgent delivery should be considered if circumstances warrant. In critical cases where disease flares are unmanageable or pose a grave risk to the mother's survival, terminating the pregnancy prematurely becomes an essential safeguard [209].

### Medication selection and safety considerations

In the clinical management of AIDs, medication selection requires careful consideration of both effective disease control and the safety of the fetus. According to the FDA's Pregnancy and Lactation Labeling Rule (PLLR) [210], drug labels must provide detailed information on medication use during pregnancy and lactation to assist healthcare professionals and patients in better assessing the risks and benefits of the drug. Based on current research and clinical data, relatively

safe medications include corticosteroids, sulfasalazine, and LMWH, all of which are frequently utilized to manage disease activity and mitigate the risk of APOs [210]. Medications requiring caution, such as azathioprine, nonsteroidal anti-inflammatory drugs, and biologic agents, call for close monitoring of maternal and fetal health when used. Prohibited medications include chlorambucil, cyclophosphamide, leflunomide, and MTX as they can lead to severe teratogenic effects [210]. Through careful selection of therapies and close surveillance, it is possible to effectively balance disease management with maternal and fetal safety, thereby offering the safest treatment approach for pregnant patients. Table 2 summarizes the pharmacokinetic characteristics of common autoimmune medications, along with their effects on the immune microenvironment, therapeutic efficacy, and safety considerations [64,117,207,211-258].

These commonly used immunosuppressive medications improve pregnancy outcomes by modulating dysregulated immune microenvironment in patients with AID. For example, corticosteroids promote a shift from proinflammatory Th1 responses to anti-inflammatory Th2 activity, rebalancing Th1/Th2 immunity in pregnancy [259]. Calcineurin inhibitors selectively suppress T cell activation by inhibiting IL-2 synthesis, modulating T cell-mediated inflammation and offering disease control in conditions like lupus nephritis [260-262]. Sulfasalazine reduces reactive oxygen species and inflammatory mediators released by phagocytes, while TNF inhibitors directly block TNF- $\alpha$  signaling, thereby reducing inflammation in both maternal-fetal interface and maternal immune system [186,263]. In contrast, intravenous immunoglobulin (IVIG) exerts broad immunomodulatory effects, including suppression of B cell activity and enhancement of regulatory pathways [264-266]. Collectively, these medications help restore immune balance within the maternal-fetal interface, mitigating inflammation-associated pregnancy complications while reducing maternal disease flares.

From a clinical standpoint, the selection of these medications during pregnancy requires careful consideration of pharmacokinetics, placental transfer, and long-term maternal and fetal safety. Physiological changes, such as increased plasma volume, altered hepatic enzyme activity, and enhanced renal clearance, may significantly affect drug distribution, metabolism, and clearance. For instance, corticosteroids and HCQ exhibit dose-dependent pharmacokinetics and increased clearance, necessitating dose adjustments to maintain therapeutic levels while minimizing adverse effects [267]. Placental transfer varies across these drugs. Certolizumab pegol shows minimal transplacental passage,

**Table 2 Common drugs for autoimmune sexually transmitted diseases during pregnancy and their safety evaluation**

Drug category	Drug name	Pregnancy indications	Pharmacokinetics and effects on the immune microenvironment	Efficacy and biological safety	References
Safe medications	Corticosteroids	SLE, RA, APS	Dose-dependent characteristics (oral clearance, distribution volume, and free drug fraction all increase with higher doses or concentrations); Inhibits Th1-mediated proinflammatory factors and enhances Th2-driven anti-inflammatory cytokine production, facilitating a shift from cellular to humoral immunity and enabling selective immunomodulation rather than generalized immunosuppression	The minimum effective dose should be maintained during pregnancy; Long-term high-dose use may increase the risk of preterm birth and low birth weight in infants	[211-216]
	HCQ	SLE, RA, SS, autoimmune, congenital heart conduction block	Pregnancy markedly changes the pharmacokinetics of HCQ, showing faster clearance, expanded distribution volume, shortened half-life, and decreased blood drug concentration; By altering the pH of cellular lysosomes, the antigen-presenting function of macrophages and the secretion of IL-1 are weakened. This pH modification also reduces lymphocyte activation; Capable of disrupting antiphospholipid immune complexes and re-establishing annexin A5 binding to the phospholipid bilayer	Lowering disease activity, the incidence of preterm birth, IUGR, and the risk of preeclampsia in pregnant SLE patients; Treatment with hydroxychloroquine during pregnancy does not elevate the risk of fetal structural malformations or other adverse outcomes; If patients report changes in vision, visual fields, color perception, etc., during medication use, they should undergo prompt ophthalmologic evaluation. Long-term medication users are advised to have regular eye examinations	[220-227]
	Calcineurin inhibitors (cyclosporine A, tacrolimus)	SLE, SS, refractory RA	Tacrolimus crosses the placenta, with fetal blood drug concentrations reaching about 71% of those in the mother; Inhibit T cell nuclear factor, thereby inhibiting IL-2 synthesis and release, and suppressing and altering T cell proliferation and differentiation	Tacrolimus is an effective adjunctive or alternative therapy to corticosteroids for managing lupus nephritis flares or maintaining stable disease during pregnancy; During use of cyclosporine and tacrolimus, monitor blood pressure, kidney function, and blood potassium levels. Be mindful of drug interactions and monitor blood drug concentrations if necessary	[228-231]
	LMWH	Primary and secondary APS	Pregnancy markedly affects LMWH pharmacokinetics, with progressively elevated clearance and increased distribution volume, resulting in decreased anti-Xa activity; By enhancing ATIII's inhibitory effects on factor Xa and thrombin, the formation of thrombosis is effectively prevented, thereby reducing the occurrence of thrombotic-related complications	Combined use of heparin and aspirin increases live birth rates in APS patients; During pregnancy, it is often necessary to use LMWH or combine it with low-dose aspirin, selecting prophylactic or therapeutic doses based on the patient's condition	[117,217-219]
	NSAID	RA, spinal arthritis, APS, SLE	Aspirin pharmacokinetics during pregnancy are influenced by obesity/BMI, leading to reduced plasma concentrations, while its pharmacodynamics demonstrate a direct link between drug levels and platelet inhibition; Total metabolite concentrations of aspirin decrease during pregnancy, while its clearance increases; Decreasing prostaglandin production modulates immune cell activity and cytokine secretion, potentially indirectly inhibiting the proliferation of Th17 cells and the generation of PGE2. This further impacts macrophage function and reduces the secretion of proinflammatory cytokines (like TNF- $\alpha$ and IL-1 $\beta$ ), thus mitigating autoimmune reactions	During the mid-pregnancy period, NSAIDs are relatively safe to use, with non-selective COX inhibitors being the first choice. The use of NSAIDs in late pregnancy significantly increases the risk of fetal ductus arteriosus premature closure and should be avoided; It is essential to use NSAIDs during pregnancy, utilizing the minimal effective dose and limiting the duration of use as much as possible	[232-235]

		(Continued)			
Drug category	Drug name	Pregnancy indications	Pharmacokinetics and effects on the immune microenvironment	Efficacy and biological safety	References
	Sulfasalazine	RA and spondylarthritis with peripheral arthritis	Following sulfasalazine use during pregnancy, its metabolites can cross the placenta, though fetal exposure to 5-aminosalicylic acid remains minimal; only trace levels of 5-ASA are found in breast milk; In the intestines, it is metabolized into 5-ASA and sulfasalazine. The former inhibits prostaglandins and neutralizes proinflammatory oxygen radicals released by phagocytes. After 12 weeks of taking this medication, activated lymphocytes in peripheral blood decrease in arthritis patients	Sulfasalazine does not increase the risk of fetal malformations or adverse pregnancy outcomes; Pregnant women taking this drug should supplement with folic acid to lower the risks of fetal cleft lip, cardiovascular defects, and urethral malformations; For breastfeeding mothers taking sulfasalazine, normal breastfeeding is safe for healthy full-term infants. However, caution is required when breastfeeding preterm infants, those with glucose-6-phosphate dehydrogenase deficiency, and infants with hyperbilirubinemia; High-dose sulfasalazine (3 g/d) during breastfeeding may cause hemorrhagic diarrhea in infants; breastfeeding or medication should be discontinued if this occurs	[207,236-240]
Selective use of safe medications	Azathioprine (AZA)	SLE, SS, etc.	Systematic changes in 6-thioguanine nucleotide (6-TGN) concentrations, a metabolite of AZA, occur during pregnancy and the perinatal period, while concentrations of 6-methylmercaptopurine nucleotide (6-MMPN) are higher during pregnancy; Neonates are only transiently exposed to low concentrations of thiopurine metabolites, which are cleared within 6 weeks without causing anemia; Interfere with the synthesis of adenine and guanine nucleotides, thereby inhibiting the synthesis and growth of activated lymphocytes	The use of AZA during pregnancy is considered safe; Closely monitor complete blood counts to early detect potential bone marrow suppression	[241-243]
	Tumor necrosis factor inhibitors (TNFi)	RA and spondylarthritis	Certolizumab pegol, lacking an Fc fragment, shows minimal placental transfer and can be continued throughout pregnancy. Other TNFi (e.g., etanercept, infliximab) contain the IgG1 Fc region and cross the placenta increasingly in late pregnancy; thus, discontinuation before the third trimester is recommended; Suppresses TNF- $\alpha$ activity, decreases proinflammatory cytokine secretion, alleviates inflammation, and mitigates damage to cartilage and bone; Limits antioxidant effects, inhibits activation of T cells and macrophages, suppresses necrosis factor production, and reduces local tissue infiltration	TNFi are effective in controlling disease activity in patients with immune-mediated inflammatory diseases, and are relatively safe for use during pregnancy, though attention should be paid to their potential side effects	[244-249]
	IVIg	Inflammatory myopathies, catastrophic APS, SLE without secondary immunocytopenias	IVIg exposure levels remain stable during pre-pregnancy, early pregnancy, and mid-pregnancy, with a dose-dependent profile; IVIg has roles in modulating lymphocyte immune function, suppressing B cell and antibody activity, blocking Fc receptors, inhibiting complement function, and reducing the activity of NK cells. Moreover, the microbe-antigen-specific binding characteristics of the immunoglobulin IgG-F(ab') <sub>2</sub> fragment can offer passive immunity to the organism	IVIg treatment can result in favorable pregnancy outcomes, yet fetal health status requires close monitoring throughout therapy, particularly in late gestation	[250,251]

(Continued)

Drug category	Drug name	Pregnancy indications	Pharmacokinetics and effects on the immune microenvironment	Efficacy and biological safety	References
Avoidance of drug use	MTX	RA, psoriatic arthritis and other forms of inflammatory arthritis	MTX inhibits dihydrofolate reductase, reducing the synthesis of tetrahydrofolate, thereby inhibiting the synthesis of pyrimidines and purines, and suppressing immune cell proliferation; Methotrexate polyglutamates inhibit 5-aminoimidazole-4-carboxamide ribonucleotide (AICAR) transformingylase, leading to AICAR accumulation and promoting adenosine release, which suppresses inflammation by binding to cell surface receptors	Discontinue medication at least 3 months before pregnancy. Women who have received MTX treatment within 3 months prior to conception should supplement with folic acid before pregnancy and throughout the entire pregnancy. MTX is contraindicated during breastfeeding	[252-254]
	Cyclophosphamide	SLE, RA, systemic necrotizing vasculitis, progressive systemic sclerosis	The metabolites of cyclophosphamide undergo addition reactions with DNA bases, leading to DNA strand breaks or cross-links, which inhibit DNA replication and transcription, ultimately suppressing cell proliferation and division, producing cytotoxic effects; Suppresses the proliferation of immune cells, especially B and T cells, reduces inflammatory responses, and effectively inhibits abnormal immune reaction	Medication should be discontinued for 6 months prior to conception	[255]
	Leflunomide	Refractory lupus nephritis	By inhibiting DHODH, it disrupts the synthesis of pyrimidine nucleotides, thereby inhibiting DNA and RNA synthesis, especially affecting immune cells that are rapidly proliferating	Discontinue the medication for 2 years before planning pregnancy (if using cholestyramine or other agents to eliminate leflunomide from the enterohepatic circulation, it is recommended to discontinue the medication for 3-6 months)	[64,256]
	MMF	Moderate to severe lupus nephritis, Rheumatic disease-associated interstitial lung disease, etc.	Inhibition of adenosine deaminase disrupts the synthesis of purine nucleotides, especially impacting the proliferation of B and T cells	Discontinuation of medication is required at least 6 weeks before planning pregnancy	[64,257]
	Tripterygium wilfordii Hook. f.	RA, Behçet's syndrome triad, autoimmune hepatitis	By inhibiting T cell proliferation and reducing cytokine release, it regulates abnormal immune responses	Discontinuation of medication is required at least 6 months before planning pregnancy	[64,258]

SLE. Systemic lupus erythematosus; RA. Rheumatoid arthritis; APS. Antiphospholipid syndrome; HCQ. Hydroxychloroquine; SS. Sjögren's syndrome; IUGR. Intrauterine growth restriction; LMWH. Low-molecular-weight heparin; NSAID. Non-steroidal anti-inflammatory drug; anti-Xa. Anti-factor Xa; ATIII. Antithrombin III; ASA. Acetylsalicylic acid; BMI. Body mass index; Th17. T helper 17; PGE2. Prostaglandin E2; TNF- $\alpha$ . Tumor necrosis factor- $\alpha$ ; IL-1 $\beta$ . Interleukin-1 $\beta$ ; COX. Cyclooxygenase; AZA. Azathioprine; TNFi. Tumor necrosis factor inhibitor; 6-TGN. 6-thioguanine nucleotide; 6-MMPN. 6-methylmercaptopurine nucleotide; IVIG. Intravenous immunoglobulin; MTX. Methotrexate; AICAR. 5-aminoimidazole-4-carboxamide ribonucleotide; DNA. Deoxyribonucleic acid; RNA. Ribonucleic acid; DHODH. Dihydroorotate dehydrogenase; MMF. Mycophenolate mofetil

making it safer throughout gestation [268]. In contrast, tacrolimus and azathioprine do cross the placenta, though fetal exposure is significantly lower [269]. Their use must be balanced with risks, including fetal growth restriction (long-term corticosteroids), ductus arteriosus closure [non-steroidal anti-inflammatory drug (NSAID) use in late pregnancy], or hematologic toxicity (high-dose sulfasalazine) [270]. Long-term studies generally support the biosafety of HCQ, certolizumab, and low-dose azathioprine, with no significant increase in congenital anomalies, though careful fetal monitoring and dose optimization are critical [269]. Overall, individualized approach is essential to ensure both maternal disease control and fetal safety.

## Conclusions and outlook

The immune microenvironment in AIDs undergoes dynamic and disease-specific changes during pregnancy, significantly influencing maternal and fetal outcomes. Compared to normal pregnancy, women with AIDs are at greater risk of APOs, largely due to the failure of the typical shift from a proinflammatory to an anti-inflammatory immune phenotype at the maternal-fetal interface. This is especially evident in SLE, APS, SS, and AITD, where reduced Treg levels and antibody-mediated interference with embryo implantation and placental development are common. Conversely, certain AIDs, including RA, MS, and CD, often show clinical improvement, potentially driven by a Th2-skewed immune response, hormonal modulation, and changes in antibody glycosylation. This heterogeneity in immune responses underscores the complexity of pregnancy-related immune adaptations across different AIDs. Moreover, the disease-specific effects of immunomodulatory therapies highlight the limitations of uniform treatment strategies, emphasizing the need for individualized management.

Despite progress, critical gaps remain, the mechanisms underlying pregnancy-induced remission in RA are still unclear, and localized immune alterations in diseases like SLE remain insufficiently characterized. These challenges hinder the development of optimized therapeutic approaches. Advances in single-cell sequencing, epigenetics, and immunomonitoring technologies now offer opportunities to dissect the immunological heterogeneity of AIDs during pregnancy. Integrating these insights into clinical care, through early biomarker identification, precise disease classification, and individualized treatment planning, will advance the management of pregnancy in AIDs. Ultimately, a personalized and dynamically adaptive approach, supported by close collaboration between rheumatologists and obstetricians, is

essential for effective management of AIDs.

## Abbreviations

$\beta$ 2GPI:  $\beta$ 2-glycoprotein I  
ACPAs: Anti-citrullinated protein antibodies  
ADCC: Antibody-dependent cellular cytotoxicity  
AID: Autoimmune disease  
AITD: Autoimmune thyroid disease  
aPL: Antiphospholipid  
APO: Adverse pregnancy outcome  
APS: Antiphospholipid syndrome  
AZA: Azathioprine  
CCL: Chemokine C-C motif ligand  
CD: Crohn's disease  
CHB: Congenital heart block  
dNK: Decidua natural killer  
dsDNA: Double-stranded DNA  
CXCL: C-X-C motif chemokine ligand  
ER: Estrogen receptor  
EVT: Extravillous trophoblast  
FOXP3: Forkhead box P3  
HCQ: Hydroxychloroquine  
HLA: Human leukocyte antigen  
HT: Hashimoto's thyroiditis  
IBD: Inflammatory bowel disease  
IFN: Interferon  
IL: Interleukin  
LIF-R: Leukemia inhibitory factor receptor  
LMWH: Low-molecular-weight heparin  
IUGR: Intrauterine growth restriction  
MMP: Matrix metalloproteinase  
MS: Multiple sclerosis  
MTX: Methotrexate  
NETs: Neutrophil extracellular traps  
NK: Natural killer  
NLE: Neonatal lupus erythematosus  
NSAID: Non-steroidal anti-inflammatory drug  
PAPS: Primary antiphospholipid syndrome  
PD-L1: Programmed death ligand 1  
PE: Preeclampsia  
PSG: Pregnancy-specific glycoprotein  
pSS: primary Sjögren's syndrome  
RA: Rheumatoid arthritis  
SAPS: Secondary antiphospholipid syndrome  
SLE: Systemic lupus erythematosus  
SS: Sjögren's syndrome  
SSA/Ro: Sjögren's syndrome-related antigen A  
SSB/La: Sjögren's syndrome-related antigen B  
TGF: Transforming growth factor  
Th: T helper  
TNF: Tumor necrosis factor  
Treg: Regulatory T cell  
UC: Ulcerative colitis

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## Authors' contributions

ZJM and JC wrote the original draft of the manuscript. CXY, SPL,

and QG contributed to visualization and investigation. ZB and YYW supervised the study. XQR and JXZ reviewed and edited the manuscript. All authors read and approved the final manuscript.

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### Availability of data and materials

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### Declarations

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#### Competing interests

The authors declare that they have no competing interests.

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#### References

1. Cao F, He YS, Sang N, Liu YC, Hu X, Ni QY, *et al.* Age-standardized incidence, prevalence, and mortality rates of autoimmune diseases in women of childbearing age from 1990 to 2019. *Autoimmun Rev.* 2023;22(11):103450.
2. Singh M, Fayaz FFA, Wang J, Wambua S, Subramanian A, Reynolds JA, *et al.* Pregnancy complications and autoimmune diseases in women: systematic review and meta-analysis. *BMC Med.* 2024;22(1):339.
3. Szukiewicz D. Reproductive immunology and pregnancy 2.0. *Int J Mol Sci.* 2024;25(10):5132.
4. Förger F, Villiger P. Immunological adaptations in pregnancy that modulate rheumatoid arthritis disease activity. *Nat Rev Rheumatol.* 2020;16(2):113-22.
5. Østensen M, Villiger PM, Förger F. Interaction of pregnancy and autoimmune rheumatic disease. *Autoimmun Rev.* 2011;11(6-7):A437-A46.
6. Vondra S, Höbner AL, Lackner AI, Raffetseder J, Mihalic ZN, Vogel A, *et al.* The human placenta shapes the phenotype of decidual macrophages. *Cell Rep.* 2023;42(1):111977.
7. Arutyunyan A, Roberts K, Troulé K, Wong FCK, Sheridan MA, Kats I, *et al.* Spatial multiomics map of trophoblast development in early pregnancy. *Nature.* 2023;616(7955):143-51.
8. Smith-Jackson K, Harrison RA. Alternative pathway activation in pregnancy, a measured amount "complements" a successful pregnancy, too much results in adverse events. *Immunol Rev.* 2023;313(1):298-319.
9. Vilotić A, Nacka-Aleksić M, Pirković A, Bojić-Trbojević Ž, Dekanski D, Jovanović Krivokuća M. IL-6 and IL-8: an overview of their roles in healthy and pathological pregnancies. *Int J Mol Sci.* 2022;23(23):14574.
10. Robertson SA, O'Connell AC, Hudson SN, Seamark RF. Granulocyte-macrophage colony-stimulating factor (GM-CSF) targets myeloid leukocytes in the uterus during the post-mating inflammatory response in mice. *J Reprod Immunol.* 2000;46(2):131-54.
11. Giaglis S, Sur Chowdhury C, Van Breda SV, Stoikou M, Tladen AN, Daoudlarian D, *et al.* Circulatory neutrophils exhibit enhanced neutrophil extracellular trap formation in early puerperium: NETs at the nexus of thrombosis and immunity. *Int J Mol Sci.* 2021; 22(24):13646.
12. Faas MM, De Vos P. Maternal monocytes in pregnancy and preeclampsia in humans and in rats. *J Reprod Immunol.* 2017;119: 91-7.
13. Koldehoff M, Cierna B, Steckel NK, Beelen DW, Elmaagacli AH. Maternal molecular features and gene profiling of monocytes during first trimester pregnancy. *J Reprod Immunol.* 2013;99(1-2): 62-8.
14. Farias-Jofre M, Romero R, Galaz J, Xu Y, Tao L, Demery-Poulos C, *et al.* Pregnancy tailors endotoxin-induced monocyte and neutrophil responses in the maternal circulation. *Inflamm Res.* 2022;71(5-6):653-68.
15. Sureshchandra S, Marshall NE, Mendoza N, Jankeel A, Zulu MZ, Messaoudi I. Functional and genomic adaptations of blood monocytes to gravid obesity during pregnancy. *iScience.* 2021;24(6):102690.
16. Faas M, Spaans F, De Vos P. Monocytes and macrophages in pregnancy and pre-eclampsia. *Front Immunol.* 2014;5:298.
17. Br VK, Sarin SK. Acute-on-chronic liver failure: terminology, mechanisms and management. *Clin Mol Hepatol.* 2023;29(3):670-89.
18. Yao RQ, Ren C, Zheng LY, Xia ZF, Yao YM. Advances in immune monitoring approaches for sepsis-induced immunosuppression. *Front Immunol.* 2022;13:891024.
19. Domínguez-Andrés J, Novakovic B, Li Y, Scicluna BP, Gresnigt MS, Arts RJW, *et al.* The itaconate pathway is a central regulatory node linking innate immune tolerance and trained immunity. *Cell Metab.* 2019;29(1):211-20.e5.
20. Zhang X, Wei H. Role of decidual natural killer cells in human pregnancy and related pregnancy complications. *Front Immunol.* 2021;12:728291.
21. Xu JQ, Zhang WY, Fu JJ, Fang XZ, Gao CG, Li C, *et al.* Viral sepsis: diagnosis, clinical features, pathogenesis, and clinical considerations. *Mil Med Res.* 2024;11(1):78.
22. Mosimann B, Wagner M, Shehata H, Poon LCY, Ford B, Nicolaides

- KH, *et al.* Natural killer cells and their activation status in normal pregnancy. *Int J Reprod Med.* 2013;2013:906813.
23. Yang FL, Zheng QL, Jin LP. Dynamic function and composition changes of immune cells during normal and pathological pregnancy at the maternal-fetal interface. *Front Immunol.* 2019; 10:2317.
  24. Weng J, Couture C, Girard S. Innate and adaptive immune systems in physiological and pathological pregnancy. *Biology.* 2023;12(3):402.
  25. Lentz LS, Stutz AJ, Meyer N, Schubert K, Karkossa I, Von Bergen M, *et al.* Human chorionic gonadotropin promotes murine Treg cells and restricts pregnancy-harmful proinflammatory Th17 responses. *Front Immunol.* 2022;13:989247.
  26. Brown MA, Su MA. An inconvenient variable: sex hormones and their impact on T cell responses. *J Immunol.* 2019;202(7):1927-33.
  27. Wang W, Sung N, Gilman-Sachs A, Kwak-Kim J. T helper (Th) cell profiles in pregnancy and recurrent pregnancy losses: Th1/Th2/Th9/Th17/Th22/Tfh cells. *Front Immunol.* 2020;11:2025.
  28. Dos Santos Fagundes I, Brendler EP, Nunes Erthal I, Eder Ribeiro RJ, Caron-Lienert RS, Machado DC, *et al.* Total Th1/Th2 cytokines profile from peripheral blood lymphocytes in normal pregnancy and preeclampsia syndrome. *Hypertens Pregnancy.* 2022;41(1):15-22.
  29. Green ES, Moldenhauer LM, Groome HM, Sharkey DJ, Chin PY, Care AS, *et al.* Regulatory T cells are paramount effectors in progesterone regulation of embryo implantation and fetal growth. *JCI Insight.* 2023;8(11):e162995.
  30. Ohkura N, Sakaguchi S. Transcriptional and epigenetic basis of Treg cell development and function: its genetic anomalies or variations in autoimmune diseases. *Cell Res.* 2020;30(6):465-74.
  31. Cai D, Tang Y, Yao X. Changes of  $\gamma\delta$  T cell subtypes during pregnancy and their influences in spontaneous abortion. *J Reprod Immunol.* 2019;131:57-62.
  32. Yalan L, Rong T, Jin X, Wei W, Bo Z, Jiang L, *et al.* Applications of single-cell sequencing in cancer research: progress and perspectives. *J Hematol Oncol.* 2021;14(1):91.
  33. Huang C, Xiang Z, Zhang Y, Li Y, Xu J, Zhang H, *et al.* NKG2D as a cell surface marker on  $\gamma\delta$ -T cells for predicting pregnancy outcomes in patients with unexplained repeated implantation failure. *Front Immunol.* 2021;12:631077.
  34. Rizzuto G. B cell responses to the placenta and fetus. *Annu Rev Pathol.* 2025;20(1):33-58.
  35. Zeng J, Lawrence WR, Yang J, Tian J, Li C, Lian W, *et al.* Association between serum uric acid and obesity in Chinese adults: a 9-year longitudinal data analysis. *BMJ Open.* 2021;11(2):e041919.
  36. Liu JC, Zeng Q, Duan YG, Yeung WSB, Li RHW, Ng EHY, *et al.* B cells: roles in physiology and pathology of pregnancy. *Front Immunol.* 2024;15:1456171.
  37. Busse M, Campe KNJ, Nowak D, Schumacher A, Plenagl S, Langwisch S, *et al.* IL-10 producing B cells rescue mouse fetuses from inflammation-driven fetal death and are able to modulate T cell immune responses. *Sci Rep.* 2019;9(1):9335.
  38. Jia Z, Wei Y, Zhang Y, Song K, Yuan J. Metabolic reprogramming and heterogeneity during the decidualization process of endometrial stromal cells. *Cell Commun Signal.* 2024;22(1):385.
  39. Hanna J, Goldman-Wohl D, Hamani Y, Avraham I, Greenfield C, Natanson-Yaron S, *et al.* Decidual NK cells regulate key developmental processes at the human fetal-maternal interface. *Nat Med.* 2006;12(9):1065-74.
  40. Yue S, Meng J. Role of decidual natural killer cells in the pathogenesis of preeclampsia. *Am J Reprod Immunol.* 2025;93(1): e70033.
  41. Zhang J, Xue M, Huang J, He S, Zhu L, Zhao X, *et al.* Deficiency of UCHL1 results in insufficient decidualization accompanied by impaired dNK modulation and eventually miscarriage. *J Transl Med.* 2024;22(1):478.
  42. Hazan AD, Smith SD, Jones RL, Whittle W, Lye SJ, Dunk CE. Vascular-leukocyte interactions: mechanisms of human decidual spiral artery remodeling *in vitro*. *Am J Pathol.* 2010;177(2):1017-30.
  43. Buckley RJ, Whitley GS, Dumitriu IE, Cartwright JE. Macrophage polarisation affects their regulation of trophoblast behaviour. *Placenta.* 2016;47:73-80.
  44. Tilburgs T, Claas FHJ, Scherjon SA. Elsevier trophoblast research award lecture: unique properties of decidual T cells and their role in immune regulation during human pregnancy. *Placenta.* 2010; 31 Suppl:S82-S6.
  45. Oravec O, Xie Y, Balogh A, Posta M, Harms C, Farkas E, *et al.* Maternal and placental galectins: key players in the feto-maternal symbiotic tango. *Semin Immunopathol.* 2025;47(1):35.
  46. Tilburgs T, Roelen DL, Van Der Mast BJ, De Groot-Swings GM, Kleijburg C, Scherjon SA, *et al.* Evidence for a selective migration of fetus-specific CD4<sup>+</sup>CD25<sup>bright</sup> regulatory T cells from the peripheral blood to the decidua in human pregnancy. *J Immunol.* 2008;180(8):5737-45.
  47. Yang X, Tian Y, Zheng L, Luu T, Kwak-Kim J. The update immune-regulatory role of pro- and anti-inflammatory cytokines in recurrent pregnancy losses. *Int J Mol Sci.* 2022;24(1):132.
  48. Burke SD, Barrette VF, Carter AL, Gravel J, Adams MA, Croy BA. Cardiovascular adaptations of pregnancy in T and B cell-deficient mice. *Biol Reprod.* 2011;85(3):605-14.
  49. Robertson SA, Moldenhauer LM, Green ES, Care AS, Hull ML. Immune determinants of endometrial receptivity: a biological perspective. *Fertil Steril.* 2022;117(6):1107-20.
  50. Green S, Politis M, Rallis KS, Saenz De Villaverde Cortabarría A, Efthymiou A, Mureanu N, *et al.* Regulatory T cells in pregnancy adverse outcomes: a systematic review and meta-analysis. *Front Immunol.* 2021;12:737862.
  51. Tilburgs T, Schonkeren D, Eikmans M, Nagtzaam NM, Datema G, Swings GM, *et al.* Human decidual tissue contains differentiated CD8<sup>+</sup> effector-memory T cells with unique properties. *J Immunol.* 2010;185(7):4470-7.
  52. Xu YY, Wang SC, Lin YK, Li DJ, Du MR. Tim-3 and PD-1 regulate CD8<sup>+</sup> T cell function to maintain early pregnancy in mice. *J Reprod Dev.* 2017;63(3):289-94.
  53. Clowse MEB, Magder LS, Petri M. The clinical utility of measuring complement and anti-dsDNA antibodies during pregnancy in patients with systemic lupus erythematosus. *J Rheumatol.* 2011; 38(6):1012-6.
  54. Buyon JP, Kim MY, Guerra MM, Laskin CA, Petri M, Lockshin MD, *et al.* Predictors of pregnancy outcomes in patients with lupus: a cohort study. *Ann Intern Med.* 2015;163(3):153-63.
  55. Østensen M, Förger F, Nelson JL, Schuhmacher A, Hebisch G, Villiger PM. Pregnancy in patients with rheumatic disease: anti-inflammatory cytokines increase in pregnancy and decrease post partum. *Ann Rheum Dis.* 2004;64(6):839-44.
  56. Dai X, Fan Y, Zhao X. Systemic lupus erythematosus: updated insights on the pathogenesis, diagnosis, prevention and therapeutics. *Signal Transduct Target Ther.* 2025;10(1):102.
  57. Gómez-Bañuelos E, Fava A, Andrade F. An update on

- autoantibodies in systemic lupus erythematosus. *Curr Opin Rheumatol*. 2023;35(2):61-7.
58. Moysidou E, Christodoulou M, Lioulios G, Stai S, Karamitsos T, Dimitroulas T, et al. Lymphocytes change their phenotype and function in systemic lupus erythematosus and lupus nephritis. *Int J Mol Sci*. 2024;25(20):10905.
59. Cajamarca-Baron J, Sanmiguel-Reyes C, Bedoya-Loaiza JE, Castañeda-Gonzalez JP, Acelas-Gonzalez GE, Molina-Giraldo S, et al. Maternal and fetal outcomes in Latin American SLE pregnancies: a systematic review and meta-analysis. *Autoimmun Rev*. 2025;24(4):103744.
60. Seyed-Kolbadi FZ, Malektojari A, Zarei MH, Keshavarz M, Gorgin K, Bonyadi M, et al. Lupus activity and pregnancy outcomes in systemic lupus erythematosus patients undergoing assisted reproductive therapy: a systematic review and meta-analysis. *Clin Rheumatol*. 2025;44(1):33-41.
61. Smyth A, Oliveira GHM, Lahr BD, Bailey KR, Norby SM, Garovic VD. A systematic review and meta-analysis of pregnancy outcomes in patients with systemic lupus erythematosus and lupus nephritis. *Clin J Am Soc Nephrol*. 2010;5(11):2060-8.
62. Lu J, Xu D, Wan Q, Chen H. Pregnancy outcomes and risk factors analysis in patients with systemic lupus erythematosus. *BMC Pregnancy Childbirth*. 2024;24(1):495.
63. Birru Talabi M, Clowse MEB. Antirheumatic medications in pregnancy and breastfeeding. *Curr Opin Rheumatol*. 2020;32(3):238-46.
64. Rüegg L, Pluma A, Hamroun S, Cecchi I, Perez-Garcia LF, Anderson PO, et al. EULAR recommendations for use of antirheumatic drugs in reproduction, pregnancy, and lactation: 2024 update. *Ann Rheum Dis*. 2025;84(6):910-26.
65. Braga A, Neves E, Guimarães J, Braga J, Vasconcelos C. The dynamics of Th17/Treg ratio in SLE patients during pregnancy. *J Reprod Immunol*. 2022;151:103622.
66. Wind M, Fierro JJ, Bloemenkamp KWM, De Leeuw K, Lely AT, Limper M, et al. Pregnancy outcome predictors in systemic lupus erythematosus: a systematic review and meta-analysis. *Lancet Rheumatol*. 2024;6(10):e667-e83.
67. Li HT, Zhang SX, Zhang JQ, Cheng T, Liu Y, Liu HQ, et al. A decreased number of circulating regulatory T cells is associated with adverse pregnancy outcomes in patients with systemic lupus erythematosus. *Immun Inflamm Dis*. 2022;10(12):e731.
68. Alanazi H, Zhang Y, Fatunbi J, Luu T, Kwak-Kim J. The impact of reproductive hormones on T cell immunity; normal and assisted reproductive cycles. *J Reprod Immunol*. 2024;165:104295.
69. Brockmann L, Tran A, Huang Y, Edwards M, Ronda C, Wang HH, et al. Intestinal microbiota-specific Th17 cells possess regulatory properties and suppress effector T cells via c-MAF and IL-10. *Immunity*. 2023;56(12):2719-35.e7.
70. Lien HJT, Pedersen TT, Jakobsen B, Flatberg A, Chawla K, Sætrum P, et al. Single-cell resolution of longitudinal blood transcriptome profiles in rheumatoid arthritis, systemic lupus erythematosus and healthy control pregnancies. *Ann Rheum Dis*. 2024;83(3):300-11.
71. Zhang P, Mo D, Zeng W, Dai H. Association between triglyceride-glucose related indices and all-cause and cardiovascular mortality among the population with cardiovascular-kidney-metabolic syndrome stage 0-3: a cohort study. *Cardiovasc Diabetol*. 2025;24(1):92.
72. Spinillo A, Bellingeri C, Cavagnoli C, Maggio ID, Riceputi G, Ruspini B, et al. Maternal and foetal placental vascular malperfusion in pregnancies with anti-phospholipid antibodies. *Rheumatology (Oxford)*. 2021;60(3):1148-57.
73. Su Y, Hong S, Zhao A. [The changes of T lymphocyte subsets and serum interleukin-2 receptor in peripheral blood of the pregnant women complicated by systemic lupus erythematosus]. *Zhonghua Fu Chan Ke Za Zhi*. 1997;32(11):671-3.
74. Jiang M, Shen N, Zhou H, Wang Y, Lin S, Wu J, et al. The enrichment of neutrophil extracellular traps impair the placentas of systemic lupus erythematosus through accumulating decidual NK cells. *Sci Rep*. 2021;11(1):6870.
75. Götestam Skorpen C, Lydersen S, Gilboe IM, Skomsvoll JF, Salvesen KÅ, Palm Ø, et al. Disease activity during pregnancy and the first year postpartum in women with systemic lupus erythematosus. *Arthritis Care Res*. 2017;69(8):1201-8.
76. Davis-Porada J, Kim MY, Guerra MM, Laskin CA, Petri M, Lockshin MD, et al. Low frequency of flares during pregnancy and postpartum in stable lupus patients. *Arthritis Res Ther*. 2020;22(1): 52.
77. Luo P, Gao FQ, Sun W, Li JY, Wang C, Zhang QY, et al. Activatable fluorescent probes for imaging and diagnosis of rheumatoid arthritis. *Mil Med Res*. 2023;10(1):31.
78. Lv J, Xu L, Mao S. Association between disease activity of rheumatoid arthritis and maternal and fetal outcomes in pregnant women: a systematic review and meta-analysis. *BMC Pregnancy Childbirth*. 2023;23(1):724.
79. Pina Vegas L, Drouin J, Weill A, Dray-Spira R. Pregnancy outcomes in women with rheumatoid arthritis: an 11-year French nationwide study. *RMD Open*. 2024;10(1):e003762.
80. Hellgren K, Secher AE, Glintborg B, Rom AL, Gudbjornsson B, Michelsen B, et al. Pregnancy outcomes in relation to disease activity and anti-rheumatic treatment strategies in women with rheumatoid arthritis: a matched cohort study from Sweden and Denmark. *Rheumatology (Oxford)*. 2022;61(9):3711-22.
81. Zanetti A, Zambon A, Scirè CA, Bortoluzzi A. Impact of rheumatoid arthritis and methotrexate on pregnancy outcomes: retrospective cohort study of the Italian Society for Rheumatology. *RMD Open*. 2022;8(2):e002412.
82. Song YJ, Cho SK, Jung YS, Jung SY, Keum J, Nam E, et al. Medication utilisation trends during pregnancy and factors influencing adverse pregnancy outcomes in patients with rheumatoid arthritis. *RMD Open*. 2024;10(1):e003739.
83. Jang S, Kwon EJ, Lee JJ. Rheumatoid arthritis: pathogenic roles of diverse immune cells. *Int J Mol Sci*. 2022;23(2):905.
84. Wehr P, Purvis H, Law SC, Thomas R. Dendritic cells, T cells and their interaction in rheumatoid arthritis. *Clin Exp Immunol*. 2019;196(1):12-27.
85. Beck F, Nguyen P, Hoffmann A, Loyal L, Thiel A, Melzer M, et al. CD4<sup>+</sup>CD8<sup>low</sup> T cell clonal expansion dependent on costimulation in patients with rheumatoid arthritis. *Arthritis Rheumatol*. 2024;76(12):1719-29.
86. Steiner G, Toes REM. Autoantibodies in rheumatoid arthritis – rheumatoid factor, anticitrullinated protein antibodies and beyond. *Curr Opin Rheumatol*. 2024;36(3):217-24.
87. Nelson JL, Hughes KA, Smith AG, Nisperos BB, Branchaud AM, Hansen JA. Maternal-fetal disparity in HLA class II alloantigens and the pregnancy-induced amelioration of rheumatoid arthritis. *N Engl J Med*. 1993;329(7):466-71.
88. De Man YA, Bakker-Jonges LE, Goorbergh CMDVD, Tillemans SPR, Hooijkaas H, Hazes JMW, et al. Women with rheumatoid arthritis negative for anti-cyclic citrullinated peptide and rheumatoid factor are more likely to improve during pregnancy, whereas

- in autoantibody-positive women autoantibody levels are not influenced by pregnancy. *Ann Rheum Dis.* 2009;69(2):420-3.
89. Hench P. The ameliorating effect of pregnancy on chronic atrophic (infectious rheumatoid) arthritis, fibrositis, and intermittent hydrarthrosis. *Proc Staff Meet Mayo Clin;* 1938;13:161-7.
90. Jethwa H, Lam S, Smith C, Giles I. Does rheumatoid arthritis really improve during pregnancy? A systematic review and meta-analysis. *J Rheumatol.* 2019;46(3):245-50.
91. Ince-Askan H, Hazes JMW, Dolhain RJEM. Identifying clinical factors associated with low disease activity and remission of rheumatoid arthritis during pregnancy. *Arthritis Care Res.* 2017; 69(9):1297-303.
92. Raine C, Austin K, Giles I. Mechanisms determining the amelioration of rheumatoid arthritis in pregnancy: a systematic review. *Semin Arthritis Rheum.* 2020;50(6):1357-69.
93. Rook GA, Steele J, Brealey R, Whyte A, Isenberg D, Sumar N, et al. Changes in IgG glycoform levels are associated with remission of arthritis during pregnancy. *J Autoimmun.* 1991;4(5):779-94.
94. Van Der Horst-Bruinsma IE, De Vries RR, De Buck PD, Van Schendel PW, Breedveld FC, Schreuder GM, et al. Influence of HLA-class II incompatibility between mother and fetus on the development and course of rheumatoid arthritis of the mother. *Ann Rheum Dis.* 1998;57(5):286-90.
95. Schwedler C, Häupl T, Kalus U, Blanchard V, Burmester GR, Poddubnyy D, et al. Hypogalactosylation of immunoglobulin G in rheumatoid arthritis: relationship to HLA-DRB1 shared epitope, anticitrullinated protein antibodies, rheumatoid factor, and correlation with inflammatory activity. *Arthritis Res Ther.* 2018; 20(1):44.
96. Camacho EM, Farragher TM, Lunt M, Verstappen SMM, Bunn D, Symmons DPM. The relationship between post-onset pregnancy and functional outcome in women with recent onset inflammatory polyarthritis: results from the Norfolk Arthritis Register. *Ann Rheum Dis.* 2010;69(10):1834-7.
97. Szabó D, Gyebrovski B, Szarka E, Auer F, Rojkovich B, Nagy G, et al. Immunoglobulin G subclass-specific glycosylation changes in rheumatoid arthritis. *Int J Mol Sci.* 2025;26(19):9626.
98. Bondt A, Hafkenscheid L, Falck D, Kuijper TM, Rombouts Y, Hazes JMW, et al. ACPA IgG galactosylation associates with disease activity in pregnant patients with rheumatoid arthritis. *Ann Rheum Dis.* 2018;77(8):1130-6.
99. Wuhler M, Selman MHJ, McDonnell LA, Kümpfel T, Derfuss T, Khademi M, et al. Pro-inflammatory pattern of IgG1 Fc glycosylation in multiple sclerosis cerebrospinal fluid. *J Neuroinflammation.* 2015;12:235.
100. Trbojević Akmačić I, Ventham NT, Theodoratou E, Vučković F, Kennedy NA, Krištić J, et al. Inflammatory bowel disease associates with proinflammatory potential of the immunoglobulin G glycome. *Inflamm Bowel Dis.* 2015;21(6):1237-47.
101. Wouters F, Maurits MP, Van Boheemen L, Verstappen M, Mankia K, Matthijssen XME, et al. Determining in which pre-arthritis stage HLA-shared epitope alleles and smoking exert their effect on the development of rheumatoid arthritis. *Ann Rheum Dis.* 2022; 81(1):48-55.
102. Saito S, Shima T, Nakashima A, Inada K, Yoshino O. Role of paternal antigen-specific Treg cells in successful implantation. *Am J Reprod Immunol.* 2016;75(3):310-6.
103. Tham M, Schlör GR, Yerly D, Mueller C, Surbek D, Villiger PM, et al. Reduced pro-inflammatory profile of  $\gamma\delta$  T cells in pregnant patients with rheumatoid arthritis. *Arthritis Res Ther.* 2016;18:26.
104. Vieira Borba V, Shoenfeld Y. Prolactin, autoimmunity, and motherhood: when should women avoid breastfeeding?. *Clin Rheumatol.* 2019;38(5):1263-70.
105. Hemon M, Giassi M, Ghaffar Y, Martin M, Roudier J, Auger I, et al. Microchimeric cells promote production of rheumatoid arthritis-specific autoantibodies. *J Autoimmun.* 2024;146:103238.
106. Goin DE, Smed MK, Pachter L, Purdom E, Nelson JL, Kjærgaard H, et al. Pregnancy-induced gene expression changes *in vivo* among women with rheumatoid arthritis: a pilot study. *Arthritis Res Ther.* 2017;19(1):104.
107. Argyriou A, Wadsworth MH, Fienman J, Gonzalez-Sanchez AC, Ghannoum S, Krishna C, et al. Innate immune cell subsets are enriched in synovial fluid of ACPA-negative rheumatoid arthritis and characterized by distinct type I IFN gene signatures. *Ann Rheum Dis.* 2025; doi: 10.1016/j.ard.2025.07.029.
108. Binignat M, Miao BY, Wibrand C, Yang MM, Rychkov D, Flynn E, et al. Single-cell RNA-seq analysis reveals cell subsets and gene signatures associated with rheumatoid arthritis disease activity. *JCI Insight.* 2024;9(16):e178499.
109. Wright M, Smed MK, Nelson JL, Olsen J, Hetland ML, Jewell NP, et al. Pre-pregnancy gene expression signatures are associated with subsequent improvement/worsening of rheumatoid arthritis during pregnancy. *Arthritis Res Ther.* 2023;25(1):191.
110. Ostensen M, Villiger PM. The remission of rheumatoid arthritis during pregnancy. *Semin Immunopathol.* 2007;29(2):185-91.
111. Bondt A, Wuhler M, Kuijper TM, Hazes JMW, Dolhain RJEM. Fab glycosylation of immunoglobulin G does not associate with improvement of rheumatoid arthritis during pregnancy. *Arthritis Res Ther.* 2016;18(1):274.
112. Bai H, Tian J. Global research landscape on antiphospholipid syndrome and systemic lupus erythematosus: Trends, collaborations, and future directions. *Autoimmun Rev.* 2024;24(1): 103696.
113. Newman TG, Knight JS. Antiphospholipid syndrome: an antibody-mediated disease with emerging therapeutic opportunities. *Arthritis Rheumatol.* 2025;77(12):1626-34.
114. Cervera R, Serrano R, Pons-Estel GJ, Cervera R, Hualde L, Shoenfeld Y, De Ramón E, et al. Morbidity and mortality in the antiphospholipid syndrome during a 10-year period: a multicentre prospective study of 1000 patients. *Ann Rheum Dis.* 2015;74(6):1011-8.
115. Liu L, Sun D. Pregnancy outcomes in patients with primary antiphospholipid syndrome: a systematic review and meta-analysis. *Medicine.* 2019;98(20):e15733.
116. Manning JE, Arachchilage DJ. Dilemmas in the diagnosis and management of antiphospholipid syndrome. *J Thromb Haemost.* 2024;22(8):2156-70.
117. Mekinian A, Alijotas-Reig J, Carrat F, Costedoat-Chalumeau N, Ruffatti A, Lazzaroni MG, et al. Refractory obstetrical antiphospholipid syndrome: features, treatment and outcome in a European multicenter retrospective study. *Autoimmun Rev.* 2017;16(7):730-4.
118. Luo Y, Jin J, Yan Y, Zhang M, Hou L, Hou Y, et al. Hemorrhage complications in obstetric antiphospholipid syndrome: risk factors and association with adverse pregnancy outcomes. *Front Immunol.* 2023;14:1145146.
119. De Carolis S, Tabacco S, Rizzo F, Giannini A, Botta A, Salvi S, et al. Antiphospholipid syndrome: an update on risk factors for pregnancy outcome. *Autoimmun Rev.* 2018;17(10):956-66.
120. Bitsadze V, Yakubova F, Khizroeva J, Lazarchuk A, Salnikova P,

- Vorobev A, et al. Catastrophic antiphospholipid syndrome. *Int J Mol Sci*. 2024;25(1):668.
121. Sammaritano LR. Antiphospholipid syndrome. *Best Pract Res Clin Rheumatol*. 2020;34(1):101463.
122. Weinstein A, Alexander RV, Zack DJ. A review of complement activation in SLE. *Curr Rheumatol Rep*. 2021;23(3):16.
123. Zen M, Tonello M, Favaro M, Del Ross T, Calligaro A, Giollo A, et al. Antiphospholipid antibody carriers and patients with quiescent antiphospholipid syndrome show persistent subclinical complement activation. *Rheumatology (Oxford)*. 2024;63(6):1733-8.
124. Grossi C, Artusi C, Meroni P, Borghi MO, Neglia L, Lonati PA, et al.  $\beta 2$  glycoprotein I participates in phagocytosis of apoptotic neurons and in vascular injury in experimental brain stroke. *J Cerebr Blood Flow Metab*. 2021;41(8):2038-53.
125. Gao R, Zeng R, Qing P, Meng C, Cheng K, Zhang S, et al. Antiphospholipid antibodies and pregnancy outcome of assisted reproductive treatment: a systematic review and meta-analysis. *Am J Reprod Immunol*. 2021;86(4):e13470.
126. Tambralli A, Gockman K, Knight JS. NETs in APS: current knowledge and future perspectives. *Curr Rheumatol Rep*. 2020; 22(10):67.
127. Manukyan G, Kriegova E, Slavik L, Mikulkova Z, Ulehlova J, Martirosyan A, et al. Antiphospholipid antibody-mediated NK cell cytotoxicity. *J Reprod Immunol*. 2023;155:103791.
128. Yan H, Li B, Su R, Gao C, Li X, Wang C. Preliminary study on the imbalance between Th17 and regulatory T cells in antiphospholipid syndrome. *Front Immunol*. 2022;13:873644.
129. Sarmiento E, Dale J, Arraya M, Gallego A, Lanio N, Navarro J, et al. CD8<sup>+</sup>DR<sup>+</sup> T-cells and C3 complement serum concentration as potential biomarkers in thrombotic antiphospholipid syndrome. *Autoimmune Dis*. 2014;2014:868652.
130. Carbone J, Gallego A, Lanio N, Navarro J, Orera M, Aguaron A, et al. Quantitative abnormalities of peripheral blood distinct T, B, and natural killer cell subsets and clinical findings in obstetric antiphospholipid syndrome. *J Rheumatol*. 2009;36(6):1217-25.
131. Nonobe M, Otani T, Yoshihara H, Goto S, Kitaori T, Nishikawa N, et al. Prognostic value of lupus anticoagulant and anti- $\beta 2$  glycoprotein I antibody in adverse pregnancy outcomes. *Arthritis Rheumatol*. 2025; doi: 10.1002/art.43341.
132. Rand JH, Wu XX, Lapinski R, Van Heerde WL, Reutelingsperger CP, Chen PP, et al. Detection of antibody-mediated reduction of annexin A5 anticoagulant activity in plasmas of patients with the antiphospholipid syndrome. *Blood*. 2004;104(9):2783-90.
133. Wolgast LR, Arslan AA, Wu XX, Beyda JN, Pengo V, Rand JH. Reduction of annexin A5 anticoagulant ratio identifies antiphospholipid antibody-positive patients with adverse clinical outcomes. *J Thromb Haemost*. 2017;15(7):1412-21.
134. Di Simone N, Meroni PL, D'asta M, Di Nicuolo F, D'Alessio MC, Caruso A. Pathogenic role of anti-beta2-glycoprotein I antibodies on human placenta: functional effects related to implantation and roles of heparin. *Hum Reprod Update*. 2007;13(2):189-96.
135. Sim BL, Daniel RS, Hong SS, Matar RH, Ganiel I, Nakanishi H, et al. Pregnancy outcomes in women with rheumatoid arthritis: a systematic review and meta-analysis. *J Clin Rheumatol*. 2023; 29(1):36-42.
136. Mao R, Zhu L, Long R, Zhou J, Wang X, Wang M, et al. A new insight on evaluation of the fertility and pregnancy outcome in patients with primary Sjögren syndrome: a propensity score matched study in multi-IVF centers. *Reprod Biol Endocrinol*. 2024; 22(1):57.
137. Skog A, Lagnefeldt L, Conner P, Wahren-Herlenius M, Sonesson SE. Outcome in 212 anti-Ro/SSA-positive pregnancies and population-based incidence of congenital heart block. *Acta Obstet Gynecol Scand*. 2016;95(1):98-105.
138. Chen B, Zhang C, Zhou M, Deng H, Xu J, Yin J, et al. CD4<sup>+</sup> T-cell metabolism in the pathogenesis of Sjogren's syndrome. *Int Immunopharmacol*. 2025;150:114320.
139. Chen Y, Luo X, Deng C, Zhao L, Gao H, Zhou J, et al. Immunometabolic alteration of CD4<sup>+</sup> T cells in the pathogenesis of primary Sjögren's syndrome. *Clin Exp Med*. 2024;24(1):163.
140. Wu G, Wu F, Wang L, Ying L, Lu W, Qian K, et al. Application of imaging mass cytometry for spatially profiling the micro-environment of salivary glands in primary Sjögren's syndrome. *Cell Death Dis*. 2025;16(1):392.
141. Xie Y, Chai M, Xing Y, Zhou P, Wei P, Hua H. miRNA let-7f-5p-encapsulated labial gland MSC-derived EVs ameliorate experimental Sjögren's syndrome by suppressing Th17 cells via targeting RORC/IL-17A signaling axis. *J Nanobiotechnol*. 2025; 23(1):228.
142. Mavragani CP, Moutsopoulos HM. Sjögren's syndrome: old and new therapeutic targets. *J Autoimmun*. 2020;110:102364.
143. Meng Q, Ma J, Cui J, Gu Y, Shan Y. Subpopulation dynamics of T and B lymphocytes in Sjögren's syndrome: implications for disease activity and treatment. *Front Immunol*. 2024;15:1468469.
144. De Carolis S, Garufi C, Garufi E, De Carolis MP, Botta A, Tabacco S, et al. Autoimmune congenital heart block: a review of biomarkers and management of pregnancy. *Front Pediatr*. 2020;8:607515.
145. Hedlund M, Thorlacius GE, Ivanchenko M, Ottosson V, Kyriakidis N, Lagnefeldt L, et al. Type I IFN system activation in newborns exposed to Ro/SSA and La/SSB autoantibodies in utero. *RMD Open*. 2020;6(1):e000989.
146. Ivanchenko M, Thorlacius GE, Hedlund M, Ottosson V, Meneghel L, Björkander S, et al. Natural killer cells and type II interferon in Ro/SSA and La/SSB autoantibody-exposed newborns at risk of congenital heart block. *Ann Rheum Dis*. 2021;80(2):194-202.
147. Bar-Or A, Li R. Cellular immunology of relapsing multiple sclerosis: interactions, checks, and balances. *Lancet Neurol*. 2021;20(6):470-83.
148. Li R, Patterson KR, Bar-Or A. Reassessing B cell contributions in multiple sclerosis. *Nat Immunol*. 2018;19(7):696-707.
149. Prapas P, Anagnostouli M. Macrophages and HLA-class II alleles in multiple sclerosis: insights in therapeutic dynamics. *Int J Mol Sci*. 2024;25(13):7354.
150. Volpe E, Battistini L, Borsellino G. Advances in T helper 17 cell biology: pathogenic role and potential therapy in multiple sclerosis. *Mediators Inflamm*. 2015;2015:475158.
151. Cencioni MT, Mattoscio M, Magliozzi R, Bar-Or A, Muraro PA. B cells in multiple sclerosis - from targeted depletion to immune reconstitution therapies. *Nat Rev Neurol*. 2021;17(7):399-414.
152. Luchtman DW, Ellwardt E, Larochele C, Zipp F. IL-17 and related cytokines involved in the pathology and immunotherapy of multiple sclerosis: current and future developments. *Cytokine Growth Factor Rev*. 2014;25(4):403-13.
153. Marcus R. What is multiple sclerosis?. *JAMA*. 2022;328(20):2078.
154. GBD 2021 Diabetes Collaborators. Global, regional, and national burden of diabetes from 1990 to 2021, with projections of prevalence to 2050: a systematic analysis for the Global Burden of Disease Study 2021. *Lancet*. 2023;402(10397):203-34.
155. Modrego PJ, Urrea MA, De Cerio LD. The effects of pregnancy on relapse rates, disability and peripartum outcomes in women with

- multiple sclerosis: a systematic review and meta-analysis. *J Comp Eff Res*. 2021;10(3):175-86.
156. Ostrem BL, Anderson A, Conway S, Healy BC, Oh J, Jacobs D, et al. Peripartum disease activity in moderately and severely disabled women with multiple sclerosis. *Mult Scler J Exp Transl Clin*. 2022; 8(2):20552173221104918.
157. Polanczyk M, Zamora A, Subramanian S, Matejuk A, Hess DL, Blankenhorn EP, et al. The protective effect of 17beta-estradiol on experimental autoimmune encephalomyelitis is mediated through estrogen receptor-alpha. *Am J Pathol*. 2003;163(4):1599-605.
158. Bodhankar S, Vandebark AA, Offner H. Oestrogen treatment of experimental autoimmune encephalomyelitis requires 17β-oestradiol-receptor-positive B cells that up-regulate PD-1 on CD4<sup>+</sup>Foxp3<sup>+</sup> regulatory T cells. *Immunology*. 2012;137(4):282-93.
159. Schubert C, Steinberg L, Peper J, Ramien C, Hellwig K, Köpke S, et al. Postpartum relapse risk in multiple sclerosis: a systematic review and meta-analysis. *J Neurol Neurosurg Psychiatry*. 2023; 94(9):718-25.
160. López C, Comabella M, Tintoré M, Sastre-Garriga J, Montalban X. Variations in chemokine receptor and cytokine expression during pregnancy in multiple sclerosis patients. *Mult Scler*. 2006;12(4): 421-7.
161. Ji L, Li T, Chen H, Yang Y, Lu E, Liu J, et al. The crucial regulatory role of type I interferon in inflammatory diseases. *Cell Biosci*. 2023;13(1):230.
162. Guo MH, Sama P, Labarre BA, Lokhande H, Balibalos J, Chu C, et al. Dissection of multiple sclerosis genetics identifies B and CD4<sup>+</sup> T cells as driver cell subsets. *Genome Biol*. 2022;23(1):127.
163. Hohlfeld R, Dornmair K, Meinl E, Wekerle H. The search for the target antigens of multiple sclerosis, part 1: autoreactive CD4<sup>+</sup> T lymphocytes as pathogenic effectors and therapeutic targets. *Lancet Neurol*. 2015;15(2):198-209.
164. Spadaro M, Martire S, Marozio L, Mastromauro D, Montanari E, Perga S, et al. Immunomodulatory effect of pregnancy on leukocyte populations in patients with multiple sclerosis: a comparison of peripheral blood and decidual placental tissue. *Front Immunol*. 2019;10:1935.
165. Qiu K, He Q, Chen X, Liu H, Deng S, Lu W. Pregnancy-related immune changes and demyelinating diseases of the central nervous system. *Front Neurol*. 2019;10:1070.
166. Grant CR, Liberal R, Mieli-Vergani G, Vergani D, Longhi MS. Regulatory T-cells in autoimmune diseases: challenges, controversies and yet-unanswered questions. *Autoimmun Rev*. 2014;14(2):105-16.
167. Viglietta V, Baecher-Allan C, Weiner HL, Hafler DA. Loss of functional suppression by CD4<sup>+</sup>CD25<sup>+</sup> regulatory T cells in patients with multiple sclerosis. *J Exp Med*. 2004;199(7):971-9.
168. Venken K, Hellings N, Broekmans T, Hensen K, Rummens JL, Stinissen P. Natural naive CD4<sup>+</sup>CD25<sup>+</sup>CD127<sup>low</sup> regulatory T cell (Treg) development and function are disturbed in multiple sclerosis patients: recovery of memory Treg homeostasis during disease progression. *J Immunol*. 2008;180(9):6411-20.
169. Airas L, Saraste M, Rinta S, Elovaara I, Huang YH, Wiendl H, et al. Immunoregulatory factors in multiple sclerosis patients during and after pregnancy: relevance of natural killer cells. *Clin Exp Immunol*. 2008;151(2):235-43.
170. Faissner S, Bongert M, Trendelenburg P, Thiel S, Yamamura T, Hellwig K, et al. Eomesodermin-expressing CD4<sup>+</sup> Th cells and association with pregnancy in multiple sclerosis. *Ther Adv Neurol Disord*. 2024;17:17562864241229321.
171. Gavin MA, Torgerson TR, Houston E, Deroos P, Ho WY, Stray-Pedersen A, et al. Single-cell analysis of normal and FOXP3-mutant human T cells: FOXP3 expression without regulatory T cell development. *Proc Natl Acad Sci U S A*. 2006;103(17):6659-64.
172. Lal G, Bromberg JS. Epigenetic mechanisms of regulation of Foxp3 expression. *Blood*. 2009;114(18):3727-35.
173. Hu W, Dolsten GA, Wang EY, Beroshvili G, Wang ZM, Ghelani AP, et al. Temporal and context-dependent requirements for the transcription factor Foxp3 expression in regulatory T cells. *Nat Immunol*. 2025;26(11):2059-73.
174. Søndergaard HB, Airas L, Christensen JR, Nielsen BR, Börnsen L, Oturai A, et al. Pregnancy-induced changes in microRNA expression in multiple sclerosis. *Front Immunol*. 2020;11:552101.
175. Higashi-Kuwata N, Jinnin M, Makino T, Fukushima S, Inoue Y, Muchemwa FC, et al. Characterization of monocyte/macrophage subsets in the skin and peripheral blood derived from patients with systemic sclerosis. *Arthritis Res Ther*. 2010;12(4):R128.
176. Svensson-Arvelund J, Mehta RB, Lindau R, Mirrasekhan E, Rodríguez-Martínez H, Berg G, et al. The human fetal placenta promotes tolerance against the semiallogeneic fetus by inducing regulatory T cells and homeostatic M2 macrophages. *J Immunol*. 2015;194(4):1534-44.
177. Martire S, Montarolo F, Spadaro M, Perga S, Sforza ML, Marozio L, et al. A first phenotypic and functional characterization of placental extracellular vesicles from women with multiple sclerosis. *Int J Mol Sci*. 2021;22(6):2875.
178. Williams JL, Gatson NN, Smith KM, Almad A, Mctigue DM, Whitacre CC. Serum exosomes in pregnancy-associated immune modulation and neuroprotection during CNS autoimmunity. *Clin Immunol*. 2013;149(2):236-43.
179. Marquez-Pedroza J, Hernández-Preciado MR, Valdivia-Tangarife ER, Alvarez-Padilla FJ, Mireles-Ramírez MA, Torres-Mendoza BM. Pregnant women with multiple sclerosis: an overview of gene expression and molecular interaction using bioinformatics analysis. *Int J Mol Sci*. 2024;25(12):6741.
180. Zenere A, Hellberg S, Papapavlou Lingehed G, Svenvik M, Mellergård J, Dahle C, et al. Prominent epigenetic and transcriptomic changes in CD4<sup>+</sup> and CD8<sup>+</sup> T cells during and after pregnancy in women with multiple sclerosis and controls. *J Neuroinflammation*. 2023;20:98.
181. Papapavlou Lingehed G, Hellberg S, Huang J, Khademi M, Kockum I, Carlsson H, et al. Plasma protein profiling reveals dynamic immunomodulatory changes in multiple sclerosis patients during pregnancy. *Front Immunol*. 2022;13:930947.
182. Gomez-Bris R, Saez A, Herrero-Fernandez B, Rius C, Sanchez-Martinez H, Gonzalez-Granado JM. CD4 T-cell subsets and the pathophysiology of inflammatory bowel disease. *Int J Mol Sci*. 2023;24(3):2696.
183. Bröms G, Granath F, Linder M, Stephansson O, Elmberg M, Kieler H. Birth outcomes in women with inflammatory bowel disease: effects of disease activity and drug exposure. *Inflamm Bowel Dis*. 2014;20(6):1091-8.
184. Wu RY, Xiao K, Hotte N, Tandon P, Elloumi Y, Ambrosio L, et al. Elevated IL-6 and IL-22 in early pregnancy are associated with worse disease course in women with inflammatory bowel disease. *Int J Mol Sci*. 2022;23(18):10281.
185. Khalili H. Risk of inflammatory bowel disease with oral contraceptives and menopausal hormone therapy: current evidence and future directions. *Drug Saf*. 2016;39(3):193-7.

186. Nielsen OH, Gubatan JM, Juhl CB, Streett SE, Maxwell C. Biologics for inflammatory bowel disease and their safety in pregnancy: a systematic review and meta-analysis. *Clin Gastroenterol Hepatol.* 2022;20(1):74-87.e3.
187. Bogović Crnčić T, Giroto N, Ilić Tomaš M, Krištofić I, Klobučar S, Batičić L, *et al.* Innate immunity in autoimmune thyroid disease during pregnancy. *Int J Mol Sci.* 2023;24(20):15442.
188. Xu Y, Chen H, Ren M, Gao Y, Sun K, Wu H, *et al.* Thyroid autoimmunity and adverse pregnancy outcomes: a multiple center retrospective study. *Front Endocrinol (Lausanne).* 2023;14:1081851.
189. Zhang QY, Ye XP, Zhou Z, Zhu CF, Li R, Fang Y, *et al.* Lymphocyte infiltration and thyrocyte destruction are driven by stromal and immune cell components in Hashimoto's thyroiditis. *Nat Commun.* 2022;13(1):775.
190. Yao Z, Guo F, Tan Y, Zhang Y, Geng Y, Yang G, *et al.* Causal relationship between inflammatory cytokines and autoimmune thyroid disease: a bidirectional two-sample Mendelian randomization analysis. *Front Immunol.* 2024;15:1334772.
191. Ludwig RJ, Vanhoorelbeke K, Leyboldt F, Kaya Z, Bieber K, Mclachlan SM, *et al.* Mechanisms of autoantibody-induced pathology. *Front Immunol.* 2017;8:603.
192. Niafar M, Samaie V, Soltani-Zangbar MS, Motavalli R, Dolati S, Danaii S, *et al.* The association of Treg and Th17 cells development factors and anti-TPO autoantibodies in patients with recurrent pregnancy loss. *BMC Res Notes.* 2023;16:302.
193. Konova E. The role of NK cells in the autoimmune thyroid disease-associated pregnancy loss. *Clin Rev Allergy Immunol.* 2010;39(3):176-84.
194. Popko K, Osińska I, Kucharska A, Demkow U. Cytometric analysis of perforin expression in NK cells, CD8<sup>+</sup>, and CD4<sup>+</sup> lymphocytes in children with autoimmune Hashimoto's thyroiditis—a preliminary study. *J Pediatr Endocrinol Metabol.* 2015;28(7-8):789-92.
195. Fan YT, Deng YB. Effects of maternal anti-Ro/La antibodies on fetal atrioventricular conduction evaluated with echocardiography: a state-of-the-art review. *Arch Gynecol Obstet.* 2025; 312(3):733-44.
196. Sparaco M, Carbone L, Landi D, Ingrassiotta Y, Di Girolamo R, Vitturi G, *et al.* Assisted reproductive technology and disease management in infertile women with multiple sclerosis. *CNS Drugs.* 2023;37(10):849-66.
197. Leavitt VM, Dobson R, Svenningsson A. Perinatal depression and anxiety in multiple sclerosis: treatable distress. *Neurology.* 2021; 96(23):1067-8.
198. Carlson AK, Ontaneda D, Rensel MR, Cohen JA, Kunchok A. Reproductive issues and multiple sclerosis: 20 questions. *Cleve Clin J Med.* 2023;90(4):235-43.
199. Sammaritano LR, Bermas BL, Chakravarty EE, Chambers C, Clowse MEB, Lockshin MD, *et al.* 2020 American College of Rheumatology Guideline for the management of reproductive health in rheumatic and musculoskeletal diseases. *Arthritis Rheumatol.* 2020;72(4):529-56.
200. Wang X, Pang K, Wang J, Zhang B, Liu Z, Lu S, *et al.* Microbiota dysbiosis in primary Sjögren's syndrome and the ameliorative effect of hydroxychloroquine. *Cell Rep.* 2022;40(11):111352.
201. Rosta K, Binder J, Kuczwar V, Horvath M, Heinzl F, Hörhager C, *et al.* Periconceptional counselling in women with autoimmune inflammatory rheumatic diseases. *J Clin Med.* 2024;13(9):2483.
202. Andreoli L, Gerardi MC, Gerosa M, Rozza D, Crisafulli F, Erra R, *et al.* Management of pregnancy in autoimmune rheumatic diseases: maternal disease course, gestational and neonatal outcomes and use of medications in the prospective Italian P-RHEUM.it study. *RMD Open.* 2024;10(2):e004091.
203. Zhang X, Liu L, Lin S, Duan X, Luo H, Wang Y, *et al.* The conditions that patients with systemic lupus erythematosus should fulfill before pregnancy to optimize outcomes: a large-scale multicenter cohort study from China. *Arthritis Res Ther.* 2025;27(1):31.
204. Götestam Skorpen C, Hoeltzenbein M, Tincani A, Fischer-Betz R, Elefant E, Chambers C, *et al.* The EULAR points to consider for use of antirheumatic drugs before pregnancy, and during pregnancy and lactation. *Ann Rheum Dis.* 2016;75(5):795-810.
205. Zhang N, Zhang HX, Li YW, Li Y. Benefits of hydroxychloroquine combined with low-dose aspirin on pregnancy outcomes and serum cytokines in pregnant women with systemic lupus erythematosus. *Drugs in R&D.* 2023;23(1):35-42.
206. Radin M, Cecchi I, Schreiber K, Rubini E, Roccatello D, Cuadrado MJ, *et al.* Pregnancy success rate and response to heparins and/or aspirin differ in women with antiphospholipid antibodies according to their Global Antiphospholipid Syndrome Score. *Semin Arthritis Rheum.* 2020;50(3):553-6.
207. Stockfelt M, Torell A, Gunnarsson I, Svenungsson E, Zickert A, Sennström MM, *et al.* Plasma interferon-alpha protein levels during pregnancy are associated with lower birth weight in systemic lupus erythematosus. *Rheumatology (Oxford).* 2024; 64(3):1469-75.
208. Stockfelt M, Larsson G, Engström H, Puttonen H, Zetterberg H, Blennow K, *et al.* Activated low-density granulocytes in peripheral and intervillous blood and neutrophil inflammation in placentas from SLE pregnancies. *Lupus Sci Med.* 2021;8(1):e000463.
209. Nakai T, Fukui S, Ozawa H, Kitada A, Okada M, Kishimoto M. Management of pregnant with rheumatoid arthritis: preconception care, pregnancy and lactation strategies, and maternal-fetal outcomes. *Best Pract Res Clin Rheumatol.* 2024; 39(1):102022.
210. Content and format of labeling for human prescription drug and biological products; requirements for pregnancy and lactation labeling. Final rule. *Fed Regist.* 2014;79(233):72063-103.
211. Ryu RJ, Easterling TR, Caritis SN, Venkataramanan R, Umans JG, Ahmed MS, *et al.* Prednisone pharmacokinetics during pregnancy and lactation. *J Clin Pharmacol.* 2018;58(9):1223-32.
212. Elenkov IJ. Glucocorticoids and the Th1/Th2 balance. *Ann N Y Acad Sci.* 2004;1024:138-46.
213. Claman HN. How corticosteroids work. *J Allergy Clin Immunol.* 1975;55(3):145-51.
214. Miljković Z, Momčilović M, Miljković D, Mostarica-Stojković M. Methylprednisolone inhibits IFN-gamma and IL-17 expression and production by cells infiltrating central nervous system in experimental autoimmune encephalomyelitis. *J Neuroinflammation.* 2009;6:37.
215. Shao S, Zhang Y, Liu J, Zeng C, Qin J, Liu Z, *et al.* Glucocorticoid use and varying doses on the long-term outcomes of offspring born to patients with systemic lupus erythematosus. *Eur J Pediatr.* 2024;183(5):2231-8.
216. Hysa E, Vojinovic T, Gotelli E, Alessandri E, Pizzorni C, Paolino S, *et al.* The dichotomy of glucocorticosteroid treatment in immune-inflammatory rheumatic diseases: an evidence-based perspective and insights from clinical practice. *Reumatologia.* 2023;61(4):283-93.
217. Lebaudy C, Hulot JS, Amoura Z, Costedoat-Chalumeau N, Serreau R, Ankri A, *et al.* Changes in enoxaparin pharmacokinetics during pregnancy and implications for antithrombotic therapeutic

- strategy. *Clin Pharmacol Ther.* 2008;84(3):370-7.
218. Ambati A, Knight JS, Zuo Y. Antiphospholipid syndrome management: a 2023 update and practical algorithm-based approach. *Curr Opin Rheumatol.* 2023;35(3):149-60.
219. Hamulyák EN, Scheres LJJ, Goddijn M, Middeldorp S. Antithrombotic therapy to prevent recurrent pregnancy loss in antiphospholipid syndrome—What is the evidence?. *J Thromb Haemost.* 2021;19(5):1174-85.
220. Balevic SJ, Weiner D, Clowse MEB, Eudy AM, Maharaj AR, Hornik CP, *et al.* Hydroxychloroquine PK and exposure-response in pregnancies with lupus: the importance of adherence for neonatal outcomes. *Lupus Sci Med.* 2022;9(1):e000602.
221. Dima A, Jurcut C, Chasset F, Felten R, Arnaud L. Hydroxychloroquine in systemic lupus erythematosus: overview of current knowledge. *Ther Adv Musculoskelet Dis.* 2022;14:1759720X211073001.
222. Wu XX, Guller S, Rand JH. Hydroxychloroquine reduces binding of antiphospholipid antibodies to syncytiotrophoblasts and restores annexin A5 expression. *Am J Obstet Gynecol.* 2011;205(6):576.e7-14.
223. Nguyen NV, Sandström A, Svenungsson E, Dominicus A, Arkema EV, Simard JF. Exposure to hydroxychloroquine in early pregnancy and incidence of pre-eclampsia and pre-term delivery in patients with systemic lupus erythematosus in Sweden: a nationwide population-based cohort study. *Lancet Rheumatol.* 2025;7(10):e687-e96.
224. Zhu Q, Wang J, Sun Q, Xie Z, Li R, Yang Z, *et al.* Effect of hydroxychloroquine on pregnancy outcome in patients with SLE: a systematic review and meta-analysis. *Lupus Sci Med.* 2024;11(2):e001239.
225. Chambers CD, Johnson DL, Xu R, Luo Y, Felix R, Fine M, *et al.* Birth outcomes in women who have taken hydroxychloroquine during pregnancy: a prospective cohort study. *Arthritis Rheumatol.* 2022;74(4):711-724.
226. Alle G, Guettrot-Imbert G, Larosa M, Murarasu A, Lazaro E, Morel N, *et al.* Hydroxychloroquine levels in pregnancy and maternal-fetal outcomes in systemic lupus erythematosus patients. *Rheumatology (Oxford).* 2025;64(3):1225-33.
227. Pawlak-Buś K, Leszczyński P. Hydroxychloroquine as an important immunomodulator: a novel insight into an old drug. *Pol Arch Intern Med.* 2024;134(1):16656.
228. Hebert MF, Zheng S, Hays K, Shen DD, Davis CL, Umans JG, *et al.* Interpreting tacrolimus concentrations during pregnancy and postpartum. *Transplantation.* 2013;95(7):908-15.
229. Peleg Y, Bomback AS, Radhakrishnan J. The evolving role of calcineurin inhibitors in treating lupus nephritis. *Clin J Am Soc Nephrol.* 2020;15(7):1066-72.
230. Tarter L, Bermas BL. Expert perspective on a clinical challenge: lupus and pregnancy. *Arthritis Rheumatol.* 2024;76(3):321-31.
231. Duan XP, Zhang CB, Wang WH, Lin DH. Role of calcineurin in regulating renal potassium (K<sup>+</sup>) excretion: mechanisms of calcineurin inhibitor-induced hyperkalemia. *Acta Physiol (Oxf).* 2024;240(8):e14189.
232. Shanmugalingam R, Wang X, Münch G, Fulcher I, Lee G, Chau K, *et al.* A pharmacokinetic assessment of optimal dosing, preparation, and chronotherapy of aspirin in pregnancy. *Am J Obstet Gynecol.* 2019;221(3):255.e1-e9.
233. Boelig RC, Kaushal G, Rochani A, Mckenzie SE, Kraft WK. Aspirin pharmacokinetics and pharmacodynamics through gestation. *Am J Obstet Gynecol.* 2024;231(3):344.e1-e16.
234. Le Duc K, Gilliot S, Baudalet JB, Mur S, Boukhris MR, Domanski O, *et al.* Case report: persistent pulmonary hypertension of the newborn and narrowing of the ductus arteriosus after topical use of non-steroidal anti-inflammatory during pregnancy. *Front Pharmacol.* 2021;12:756056.
235. Arfeen M, Srivastava A, Srivastava N, Khan RA, Almahmoud SA, Mohammed HA. Design, classification, and adverse effects of NSAIDs: a review on recent advancements. *Bioorg Med Chem.* 2024;112:117899.
236. Ambrosius Christensen L, Rasmussen SN, Hansen SH, Bondesen S, Hvidberg EF. Salazosulfapyridine and metabolites in fetal and maternal body fluids with special reference to 5-aminosalicylic acid. *Acta Obstet Gynecol Scand.* 1987;66(5):433-5.
237. Macmullan PA, Madigan AM, Paul N, Peace AJ, Alagha A, Nolan KB, *et al.* Sulfasalazine and its metabolites inhibit platelet function in patients with inflammatory arthritis. *Clin Rheumatol.* 2016;35(2):447-55.
238. Ratajczak AE, Szymczak-Tomczak A, Rychter AM, Zawada A, Dobrowolska A, Kreła-Kaźmierczak I. Does folic acid protect patients with inflammatory bowel disease from complications?. *Nutrients.* 2021;13(11):4036.
239. Bröms G, Granath F, Stephansson O, Kieler H. Preterm birth in women with inflammatory bowel disease - the association with disease activity and drug treatment. *Scand J Gastroenterol.* 2016;51(12):1462-9.
240. Märginean CO, Meliț LE, Mocanu S, Märginean MO. Inflammatory bowel diseases: a burden in pediatrics: case series and a review of the literature. *Medicine.* 2017;96(11):e6329.
241. Balevic S, Sims CA, Eudy A, Smith V, Clowse M. Azathioprine metabolite levels and outcomes during pregnancies with rheumatic disease. *Lupus Sci Med.* 2024;11(1):e001036.
242. Vögelin M, Biedermann L, Frei P, Vavricka SR, Scharl S, Zeitz J, *et al.* The impact of azathioprine-associated lymphopenia on the onset of opportunistic infections in patients with inflammatory bowel disease. *PLoS One.* 2016;11(5):e0155218.
243. Mahadevan U, Long MD, Kane SV, Roy A, Dubinsky MC, Sands BE, *et al.* Pregnancy and neonatal outcomes after fetal exposure to biologics and thiopurines among women with inflammatory bowel disease. *Gastroenterology.* 2021;160(4):1131-9.
244. Richter F, Seifert O, Herrmann A, Pfüzenmaier K, Kontermann RE. Improved monovalent TNF receptor 1-selective inhibitor with novel heterodimerizing Fc. *MAbs.* 2019;11(4):653-65.
245. Cozzi G, Scagnellato L, Lorenzin M, Savarino E, Zingone F, Ometto F, *et al.* Spondyloarthritis with inflammatory bowel disease: the latest on biologic and targeted therapies. *Nat Rev Rheumatol.* 2023;19(8):503-18.
246. Syversen SW, Jørgensen KK, Goll GL, Brun MK, Sandanger Ø, Bjørlykke KH, *et al.* Effect of therapeutic drug monitoring vs. standard therapy during maintenance infliximab therapy on disease control in patients with immune-mediated inflammatory diseases: a randomized clinical trial. *JAMA.* 2021;326(23):2375-84.
247. De Felice KM, Kane S. Safety of anti-TNF agents in pregnancy. *J Allergy Clin Immunol.* 2021;148(3):661-7.
248. Smeele HTW, Röder E, Mulders AGMJ, Steegers EaP, Dolhain RJE. Tumour necrosis factor inhibitor use during pregnancy is associated with increased birth weight of rheumatoid arthritis patients' offspring. *Ann Rheum Dis.* 2022;81(10):1367-73.
249. Barenbrug L, Groen MT, Hoentjen F, Van Drongelen J, Reek JMPaVD, Joosten I, *et al.* Pregnancy and neonatal outcomes in women with immune mediated inflammatory diseases exposed

- to anti-tumor necrosis factor- $\alpha$  during pregnancy: a systemic review and meta-analysis. *J Autoimmun.* 2021;122:102676.
250. Ensom MHH, Stephenson MD. A two-center study on the pharmacokinetics of intravenous immunoglobulin before and during pregnancy in healthy women with poor obstetrical histories. *Hum Reprod.* 2011;26(9):2283-8.
251. Achiron A, Kishner I, Dolev M, Stern Y, Dulitzky M, Schiff E, *et al.* Effect of intravenous immunoglobulin treatment on pregnancy and postpartum-related relapses in multiple sclerosis. *J Neurol.* 2004;251(9):1133-7.
252. Zhao Z, Hua Z, Luo X, Li Y, Yu L, Li M, *et al.* Application and pharmacological mechanism of methotrexate in rheumatoid arthritis. *Biomed Pharmacother.* 2022;150:113074.
253. Bielack SS, Soussain C, Fox CP, Houillier C, Murciano T, Osborne W, *et al.* A European consensus recommendation on the management of delayed methotrexate elimination: supportive measures, leucovorin rescue and glucarpidase treatment. *J Cancer Res Clin Oncol.* 2024;150(10):441.
254. Cronstein BN, Aune TM. Methotrexate and its mechanisms of action in inflammatory arthritis. *Nat Rev Rheumatol.* 2020;16(3):145-54.
255. Li J, Zhang R, Chen LJ, Qu XY, Lu H, Li JY, *et al.* [Comparison of etoposide combined with G-CSF and cyclophosphamide combined with G-CSF in mobilization of autologous peripheral hematopoietic stem cells in patients with newly diagnosed multiple myeloma]. *Zhonghua Xue Ye Xue Za Zhi.* 2022;43(9):781-4. (Chinese)
256. Fox RI, Herrmann ML, Frangou CG, Wahl GM, Morris RE, Strand V, *et al.* Mechanism of action for leflunomide in rheumatoid arthritis. *Clin Immunol.* 1999;93(3):198-208.
257. Sollinger HW. Mycophenolates in transplantation. *Clin Transplant.* 2004;18(5):485-92.
258. Song CY, Xu YG, Lu YQ. Use of tripterygium wilfordii Hook F for immune-mediated inflammatory diseases: progress and future prospects. *J Zhejiang Uni Sci.* 2020;21(4):280-90.
259. Miyaura H, Iwata M. Direct and indirect inhibition of Th1 development by progesterone and glucocorticoids. *J Immunol.* 2002;168(3):1087-94.
260. Jiang Y, Tao M, Chen J, Luo L, You Q, Wu H, *et al.* Calcineurin inhibitors in the treatment of systemic lupus erythematosus during pregnancy: a narrative review with emphasis on efficacy and safety. *Eur J Obstet Gynecol Reprod Biol.* 2024;294:148-55.
261. Vasi İ, Yıldırım D, Kardaş RC, Kaya B, Duran R, Alp GT, *et al.* Calcineurin inhibitors in unplanned pregnancies with active lupus disease: a retrospective observational study. *Int J Clin Pharmacol Ther.* 2024;62(7):326-33.
262. Azizi R, Ahmadi M, Danaii S, Abdollahi-Fard S, Mosapour P, Eghbal-Fard S, *et al.* Cyclosporine A improves pregnancy outcomes in women with recurrent pregnancy loss and elevated Th1/Th2 ratio. *J Cell Physiol.* 2019;234(10):19039-47.
263. Romanowska-Próchnicka K, Felis-Giemza A, Olesińska M, Wojdasiewicz P, Paradowska-Gorycka A, Szukiewicz D. The role of TNF- $\alpha$  and anti-TNF- $\alpha$  agents during preconception, pregnancy, and breastfeeding. *Int J Mol Sci.* 2021;22(6):2922.
264. Ahmadi M, Ghaebi M, Abdolmohammadi-Vahid S, Abbaspour-Aghdam S, Hamdi K, Abdollahi-Fard S, *et al.* NK cell frequency and cytotoxicity in correlation to pregnancy outcome and response to IVIG therapy among women with recurrent pregnancy loss. *J Cell Physiol.* 2018;234(6):9428-37.
265. Ahmadi M, Abdolmohammadi-Vahid S, Ghaebi M, Aghebati-Maleki L, Afkham A, Danaii S, *et al.* Effect of Intravenous immunoglobulin on Th1 and Th2 lymphocytes and improvement of pregnancy outcome in recurrent pregnancy loss (RPL). *Biomed Pharmacother.* 2017;92:1095-102.
266. Han JW, Park JS, Kim JS, Lee SK. Efficacy of intravenous immunoglobulin in recurrent pregnancy loss: a retrospective analysis of patients with abnormal cellular immunity. *Front Endocrinol (Lausanne).* 2025;16:1546602.
267. Kemp MW, Newnham JP, Challis JG, Jobe AH, Stock SJ. The clinical use of corticosteroids in pregnancy. *Hum Reprod Update.* 2015;22(2):240-59.
268. Ion A, Dorobanțu AM, Popa LG, Mihai MM, Orzan OA. Risks of biologic therapy and the importance of multidisciplinary approach for an accurate management of patients with moderate-severe psoriasis and concomitant diseases. *Biology.* 2022;11(6):808.
269. Saad AF, Pacheco LD, Saade GR. Immunosuppressant medications in pregnancy. *Obstet Gynecol.* 2024;143(4):e94-e106.
270. Hultzsch S, Schaefer C. [Analgesic drugs during pregnancy]. *Schmerz.* 2016;30(6):583-93. (German)

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